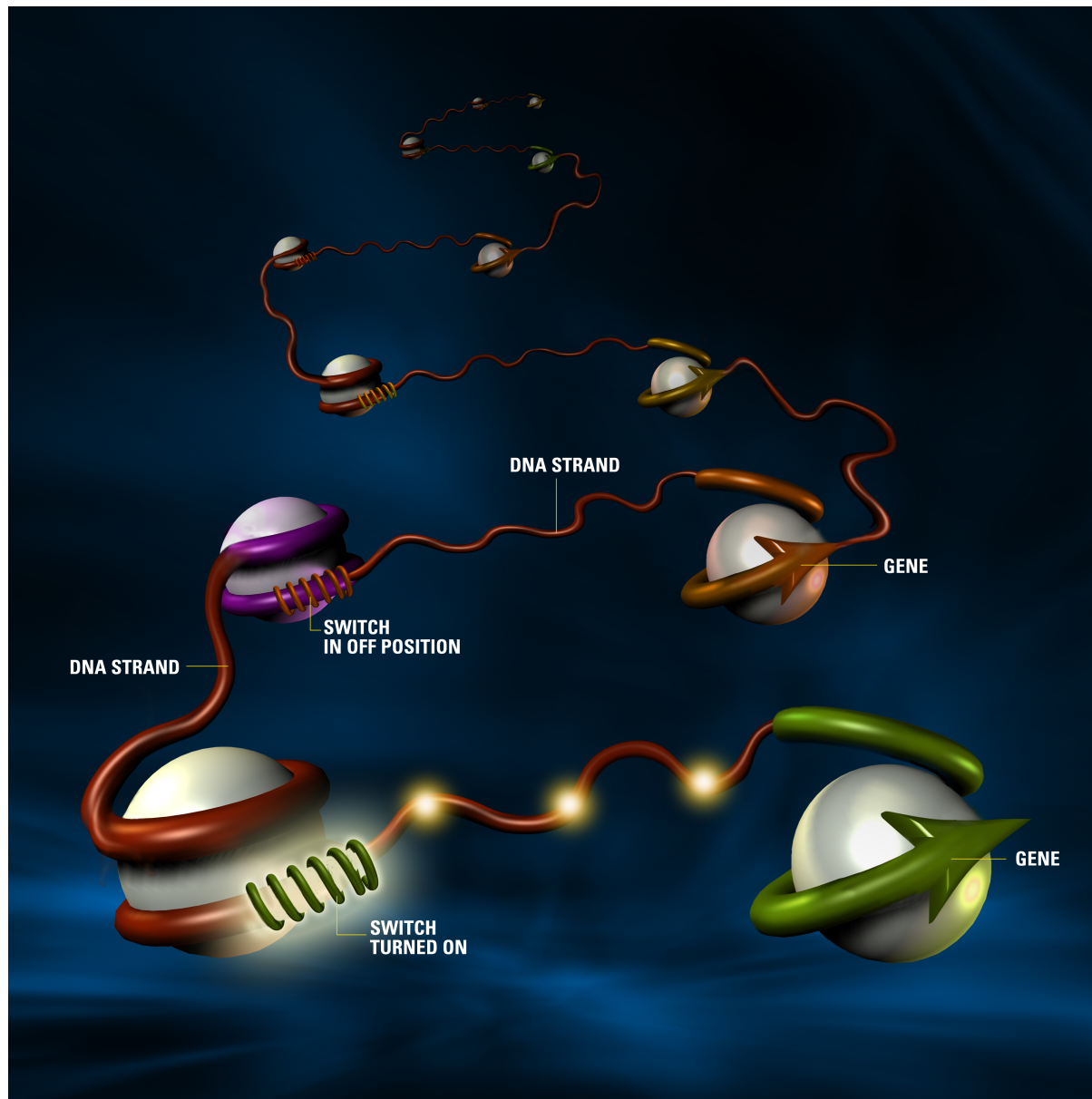
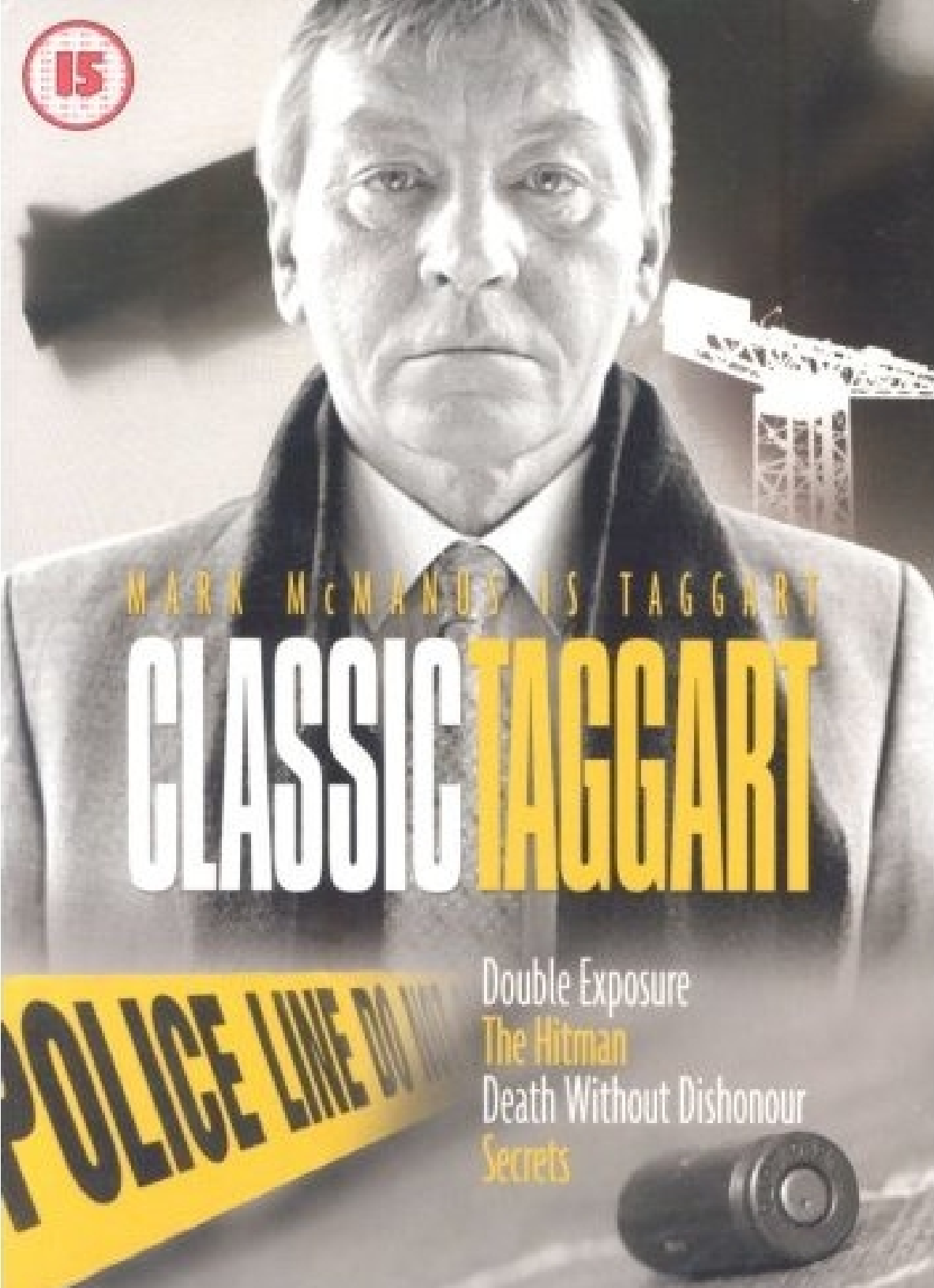


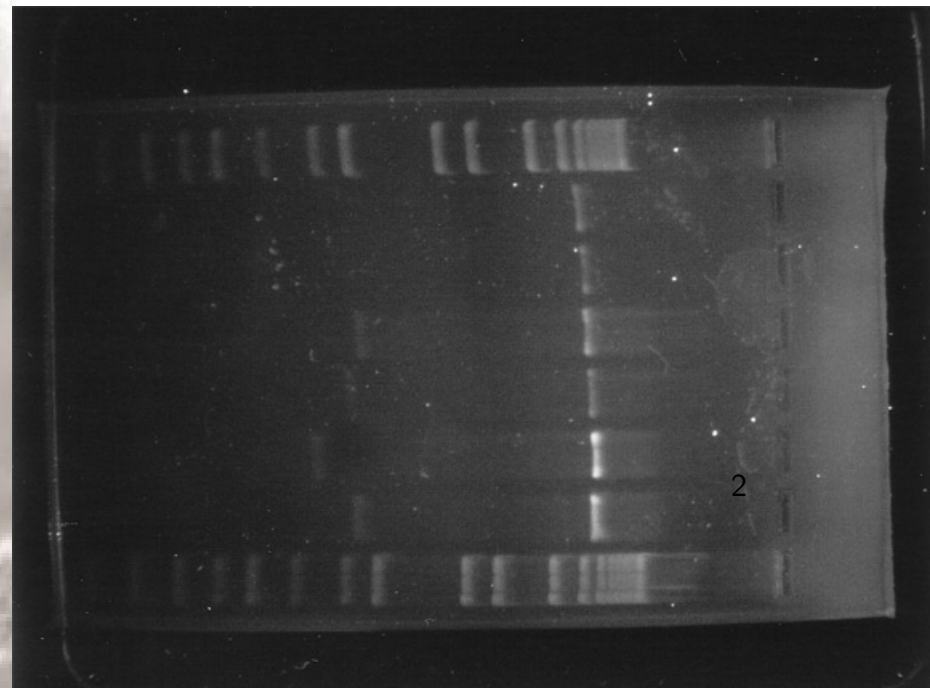
# Biological Physics of DNA



15

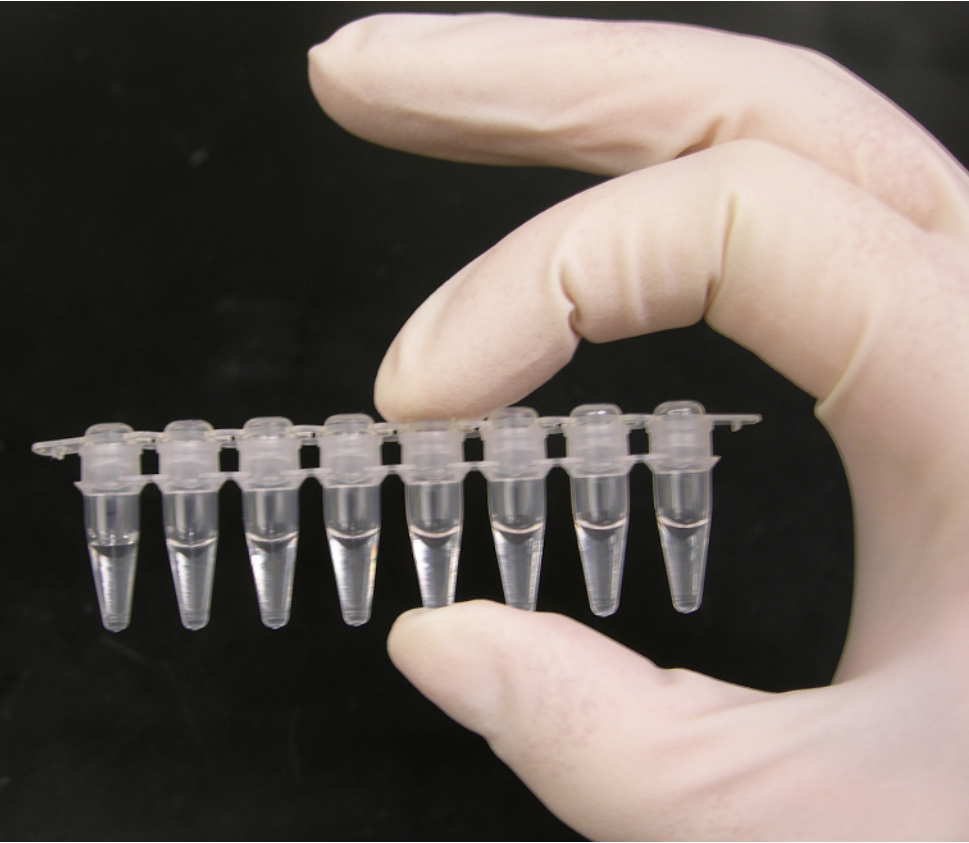


# Whodunnit...

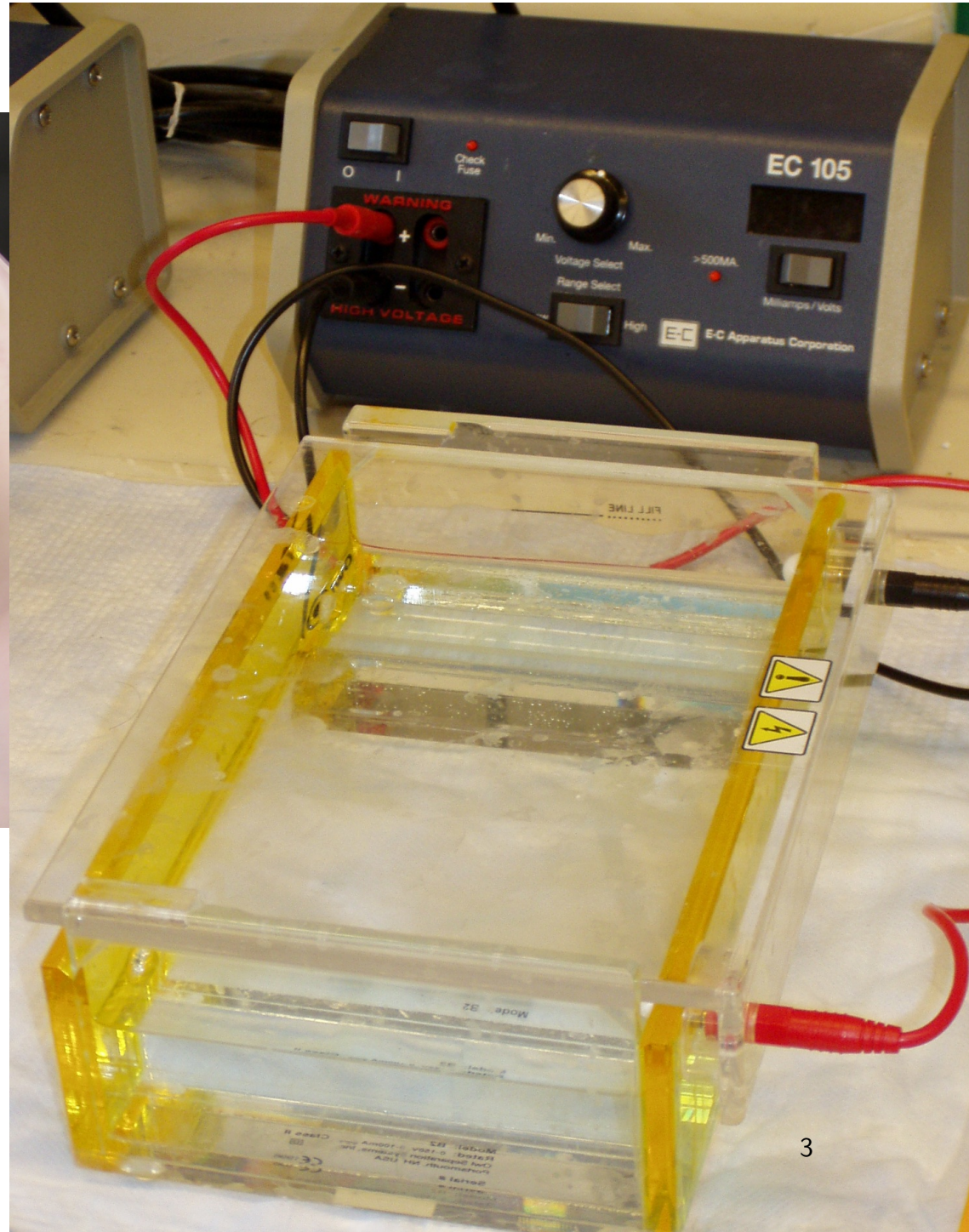




# DNA multiplication/PCR



# Gel electrophoresis



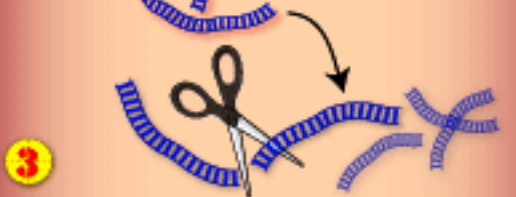


# FORENSIC DNA ANALYSIS

**1** Collect evidence from the crime scene



**2** Isolate DNA from an evidence sample



**3** Cut the DNA into fragments using specialized protein "scissors" called restriction enzymes. For every person, the sizes of the cut fragments are unique - except for identical twins.

**4** Separate the negatively charged DNA fragments in a gel by passing an electric current through it.



**6** Probe the membrane with DNA fragments that complement the DNA sequence of the fragments of interest

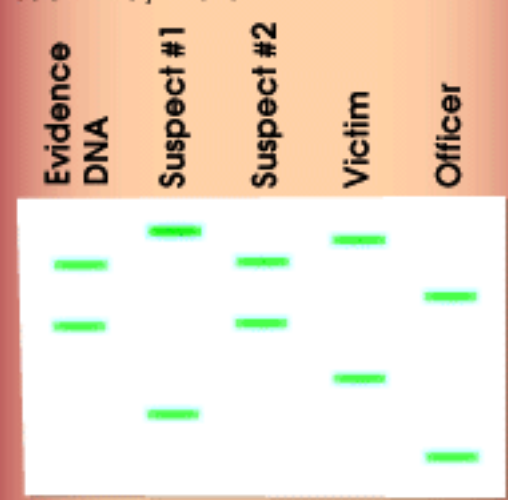


electric current through it.

**5** Transfer the DNA fragments from the gel to a sheet of membrane

**6** Probe the membrane with DNA fragments that complement the DNA sequence of the fragments of interest.

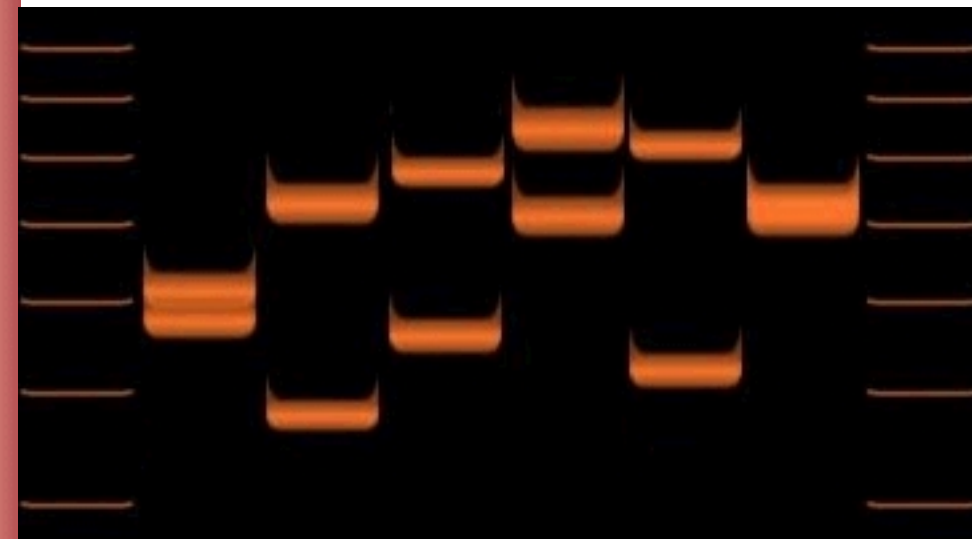
**7** Compare the fragment profile of the evidence DNA with those of the suspects, detective and victim to see if they match.



**8** Re-probe the membrane up to 10 more times to identify different fragments.

If the profiles from the evidence DNA and a suspect match multiple times, then it is very likely that the evidence DNA came from the suspect.

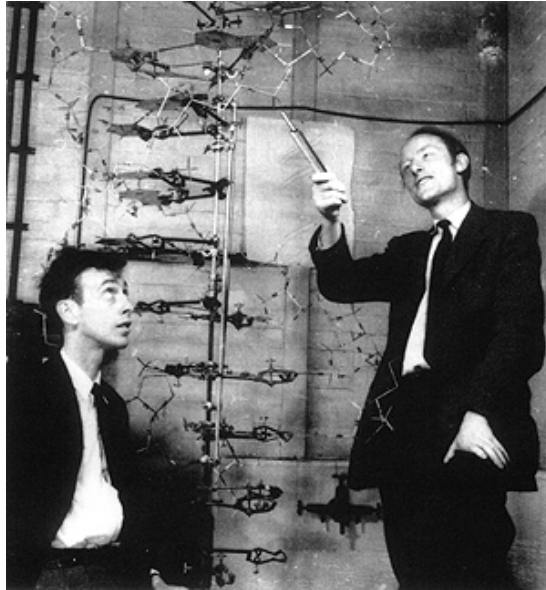
NB: betw 4 & 5 denature fragments to ssDNA  
 Labelling: eg radioact probe fragm & Xray film



Variations of VNTR allele lengths in 6 individuals



# Chief character: DeoxyriboNucleic Acid



$$d \simeq 2\text{nm}$$

$$\Delta_{\text{bp}-\text{bp}} \simeq 3.5\text{\AA}$$

$$l_p(\text{dsDNA}) \simeq 50\text{nm}$$

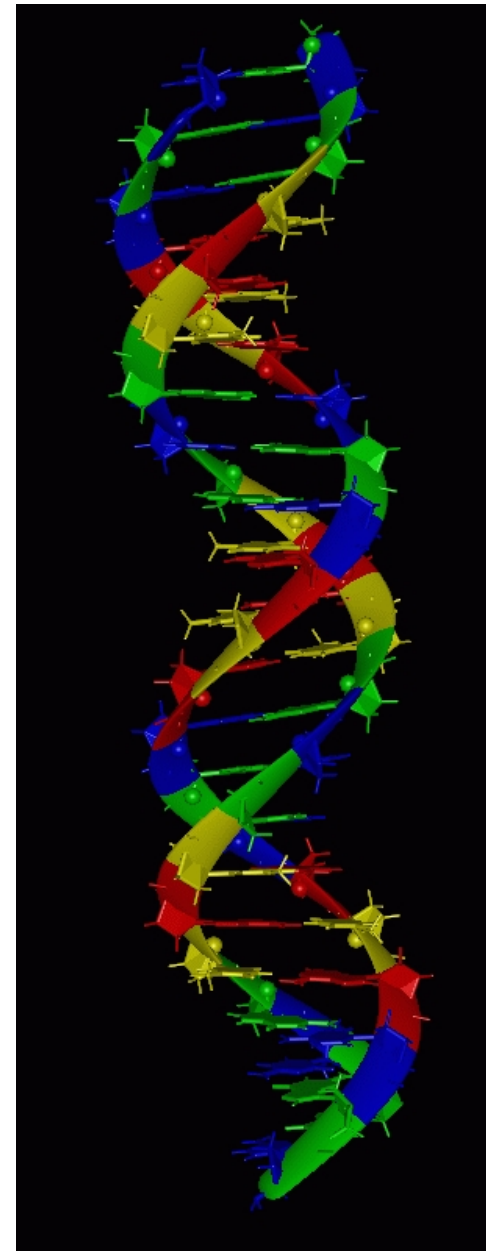
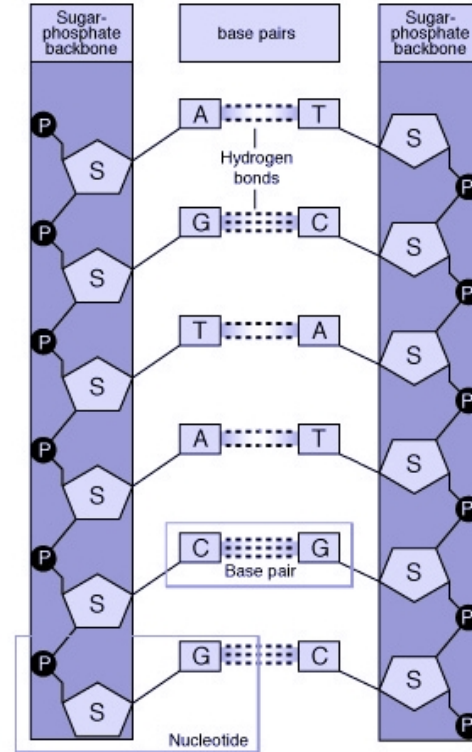
$$l_p(\text{ssDNA}) \simeq 1\text{nm}$$

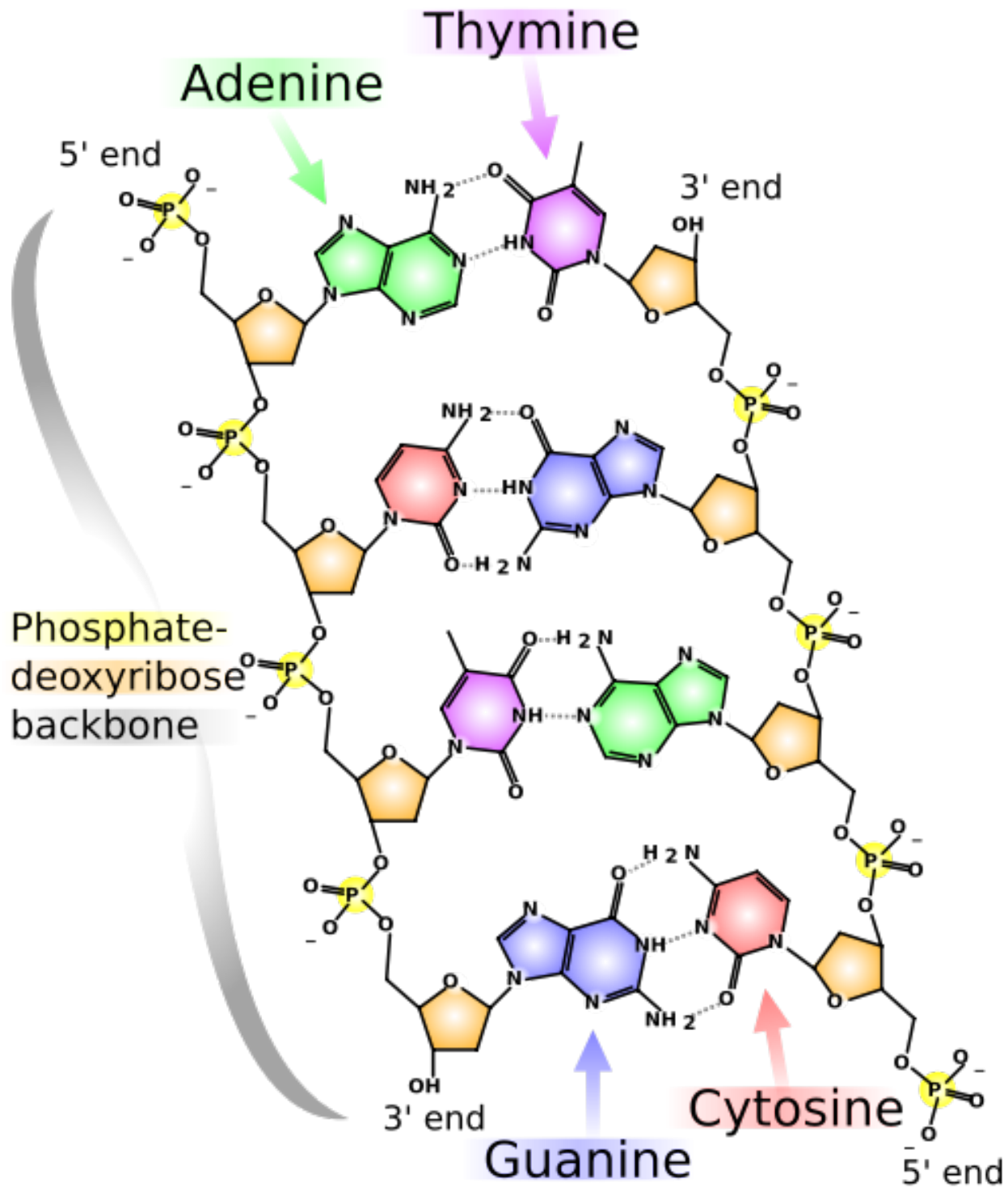
$\phi 29$ -phage	$6\mu\text{m}$
------------------	----------------

E.coli	$3\text{mm}$
--------	--------------

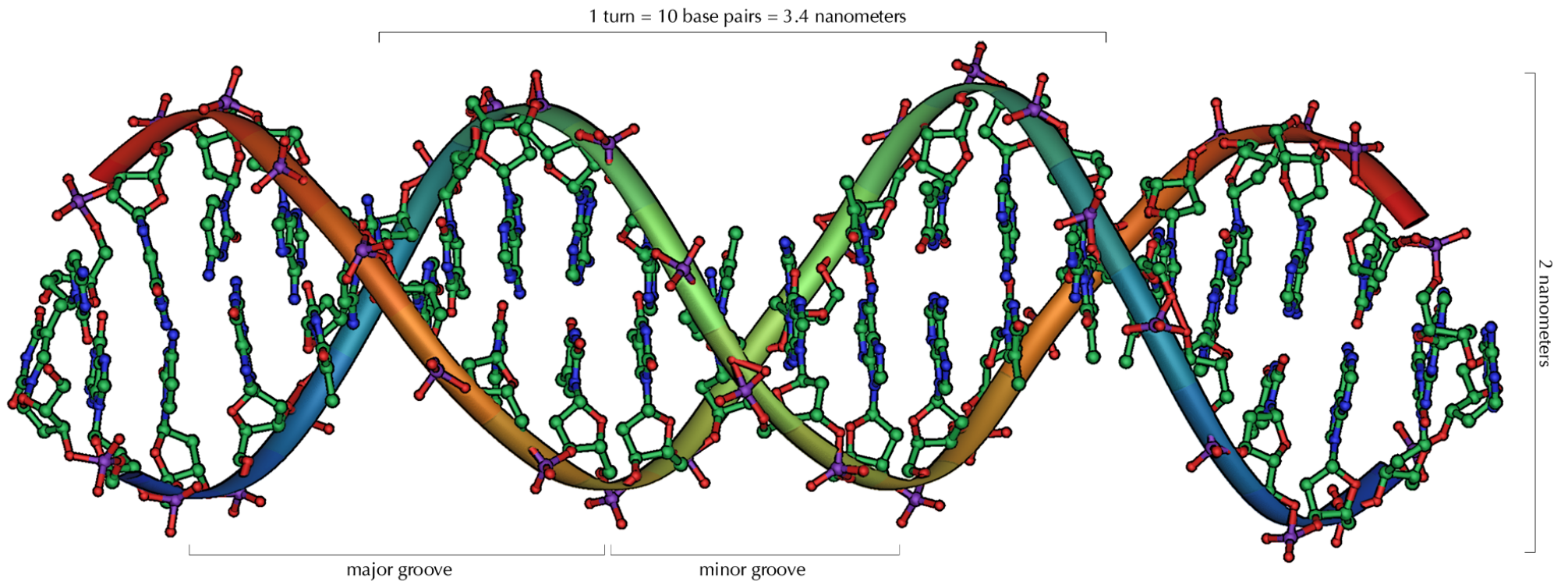
Human cell	$2\text{m}$
------------	-------------

Sth Amer lungfish	$35\text{m}$
-------------------	--------------

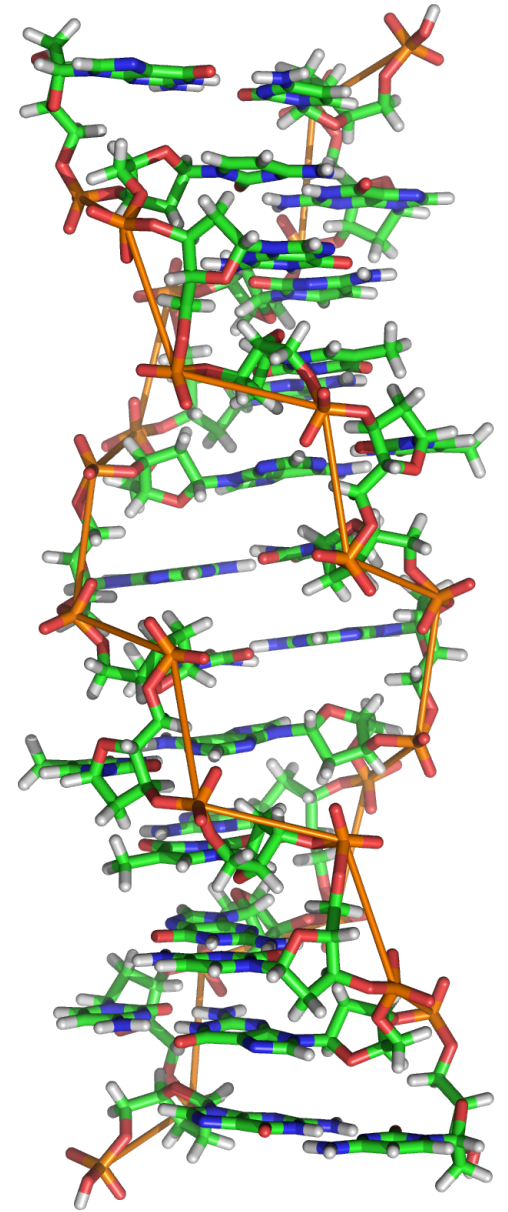
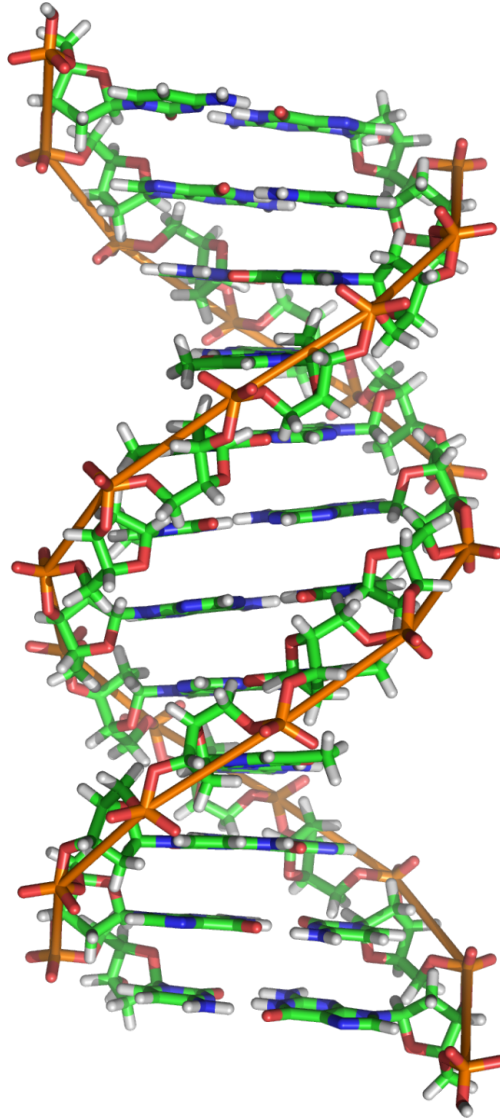
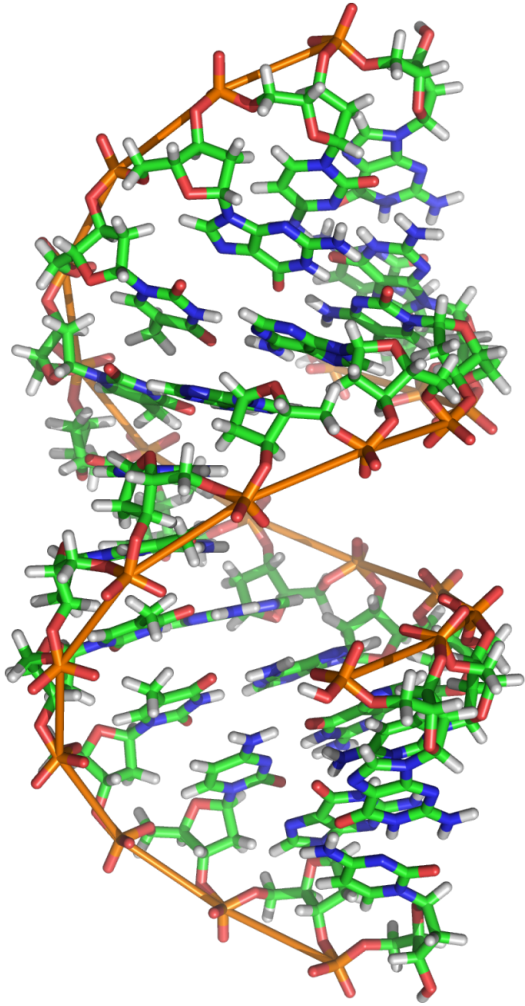








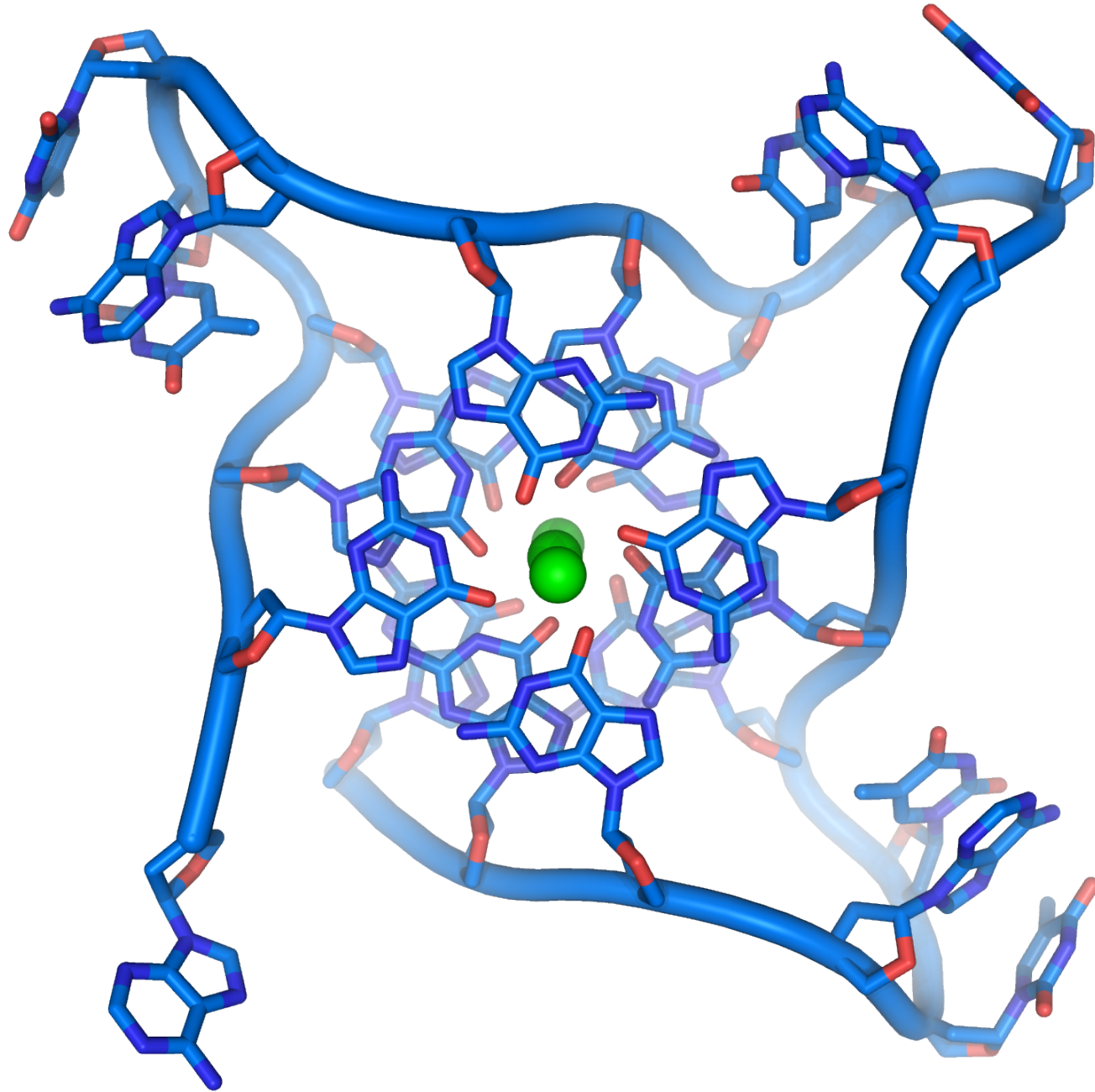
# A-DNA, B-DNA, Z-DNA



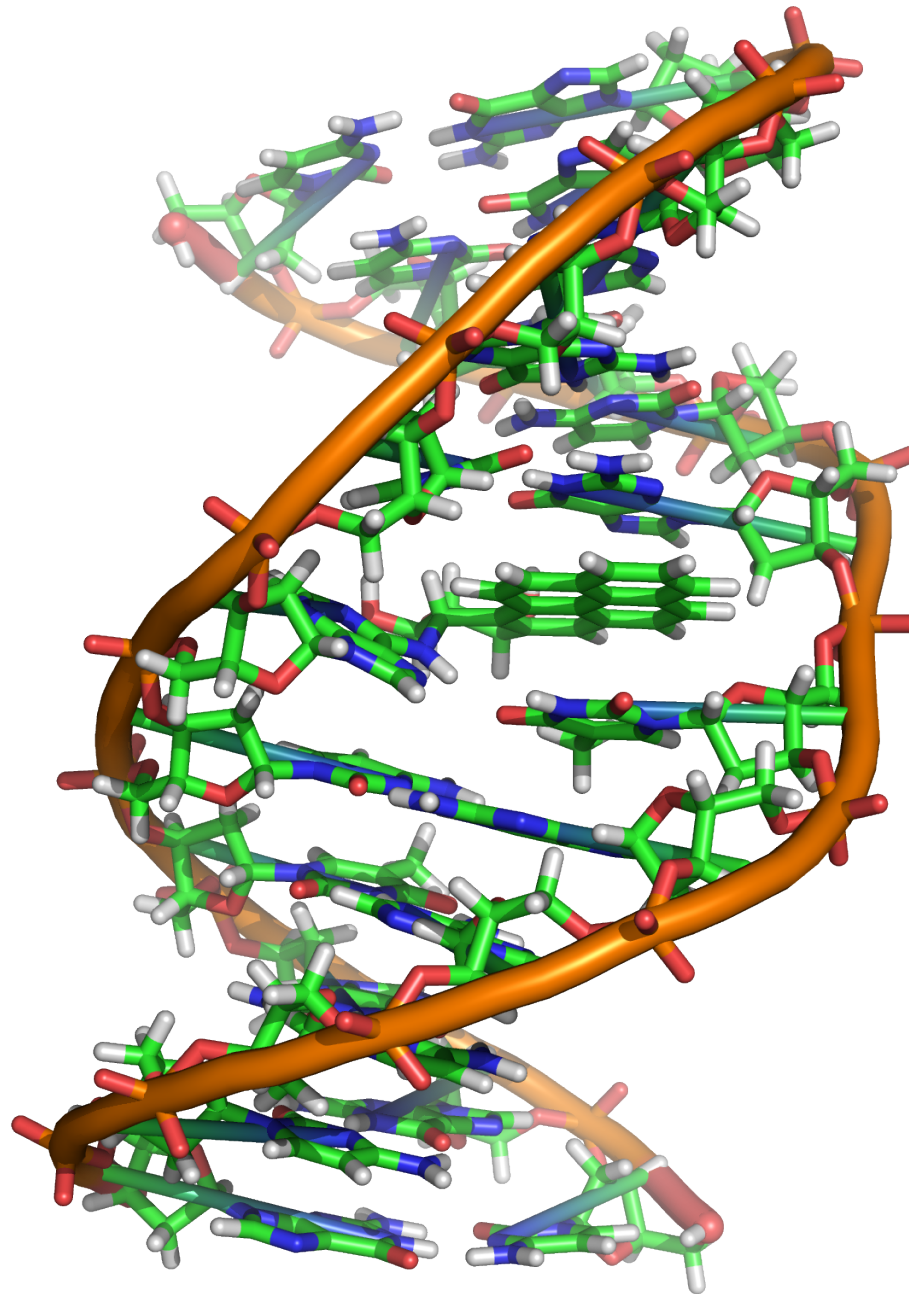


# Telomeric quadruplex DNA

GC-rich, alternative to telomere loop

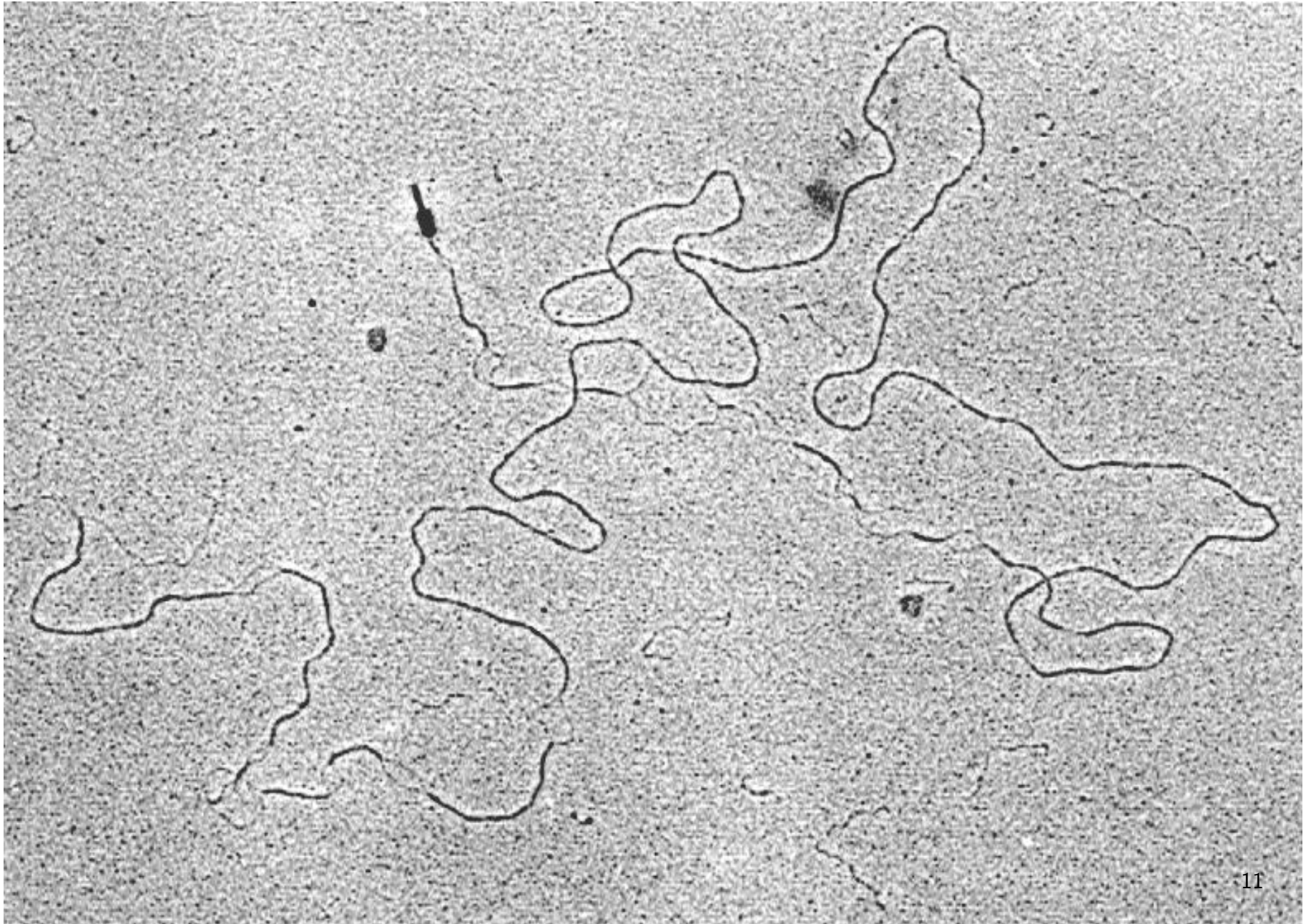


# DNA mutation induced by tobacco benzopyrene





# Packaging of DNA in bacteria



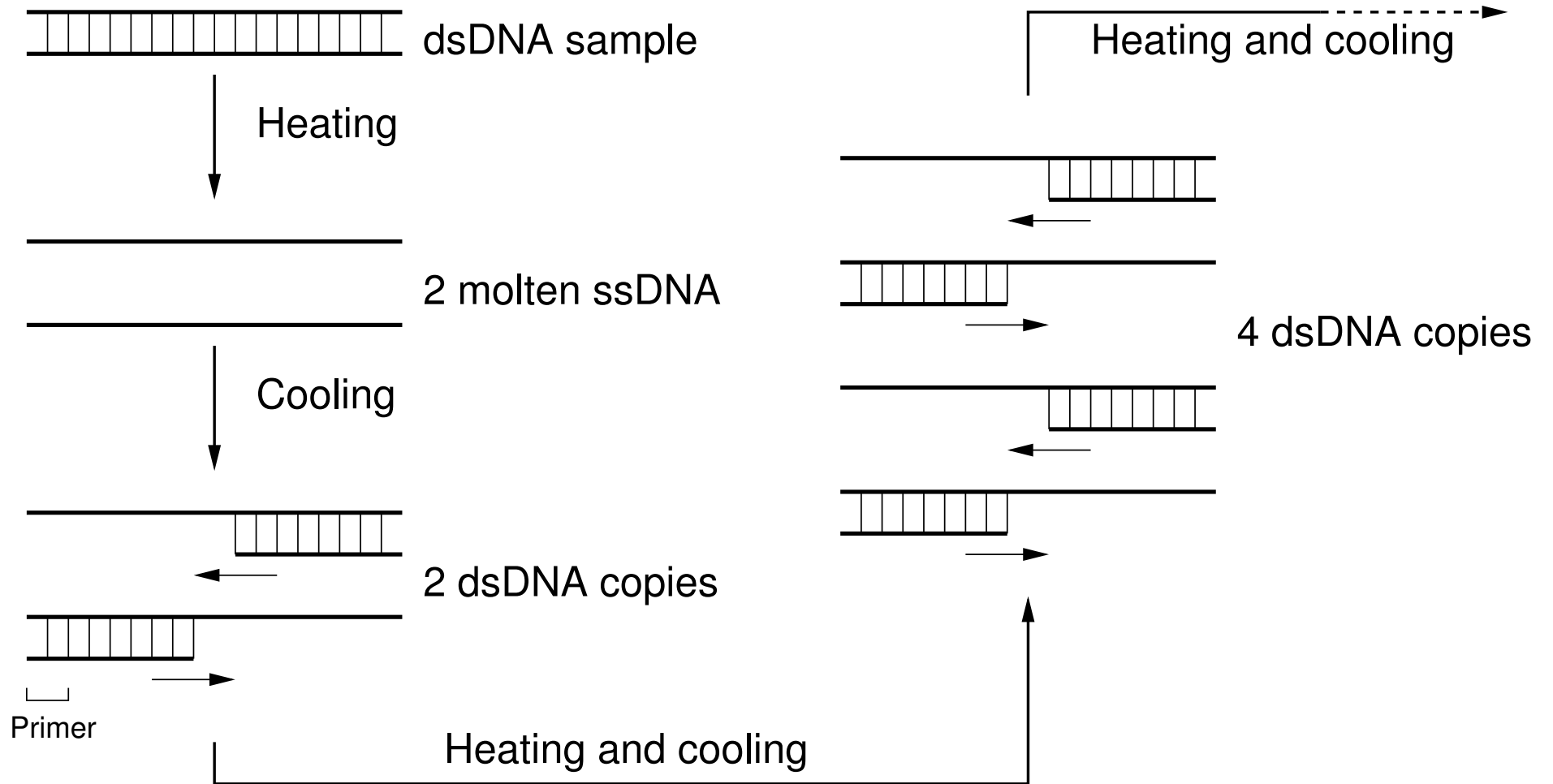


# DNA melting



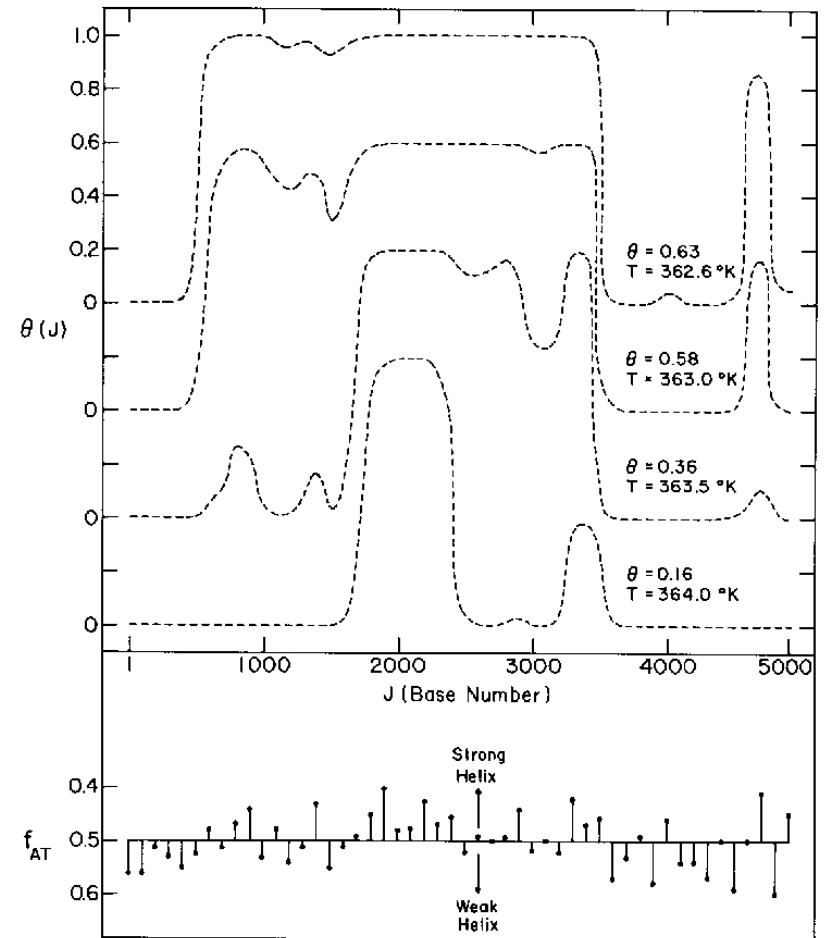
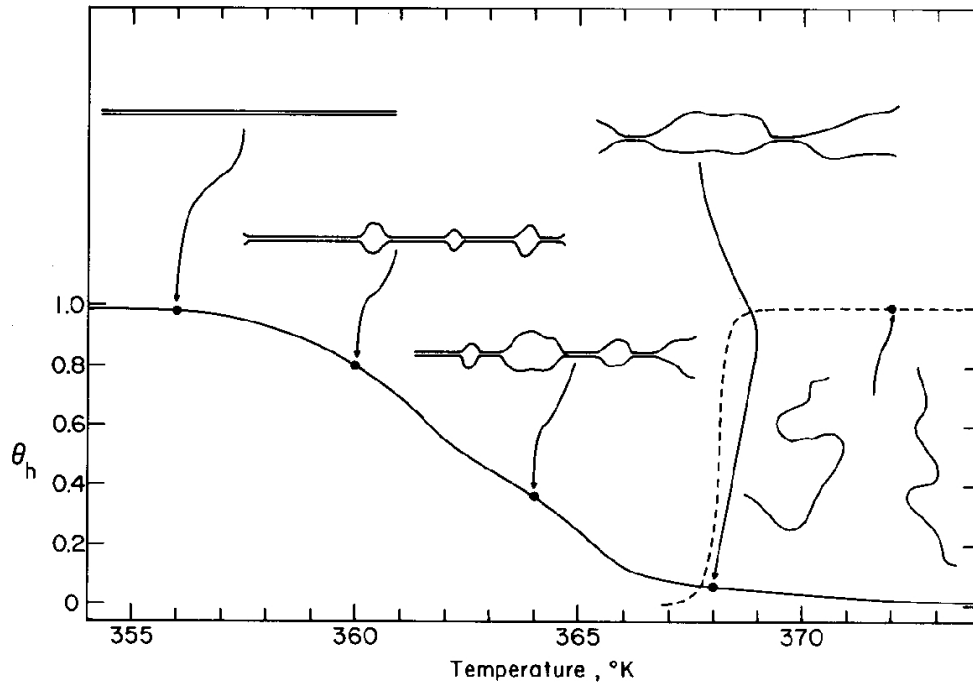


# Polymerase chain reaction



# DNA melting in bulk solution (UV absorption)

Thermal melting profile:

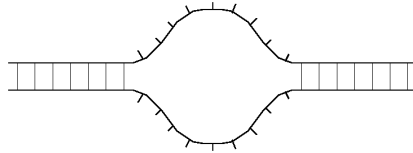




# DNA stability and function

Partition factor for bubble of  $m$  broken bps:

$$\mathcal{L}(m) = \sigma_0 u^m (1 + m)^{-c}$$



$$u = \exp\left(\beta[\Delta G_{ij} + \mathfrak{T}\theta_0]\right) \therefore \theta_0 = \frac{2\pi}{10.35}$$

$$\begin{array}{l} \cdot\text{AA}\cdot \\ \cdot\text{TT}\cdot \end{array} \Delta G = -8.45 \frac{\text{kcal}}{\text{mol}} + \underbrace{24.86 \frac{\text{cal}}{\text{mol} \cdot \text{K}} T}_{7.72 \text{kcal/mol@}37^\circ\text{C}}$$

$$\begin{array}{l} \cdot\text{AA}\cdot \\ \cdot\text{TT}\cdot \end{array} T_m \approx 68^\circ\text{C} \wedge \begin{array}{l} \cdot\text{GG}\cdot \\ \cdot\text{CC}\cdot \end{array} T_m \approx 102^\circ\text{C}$$

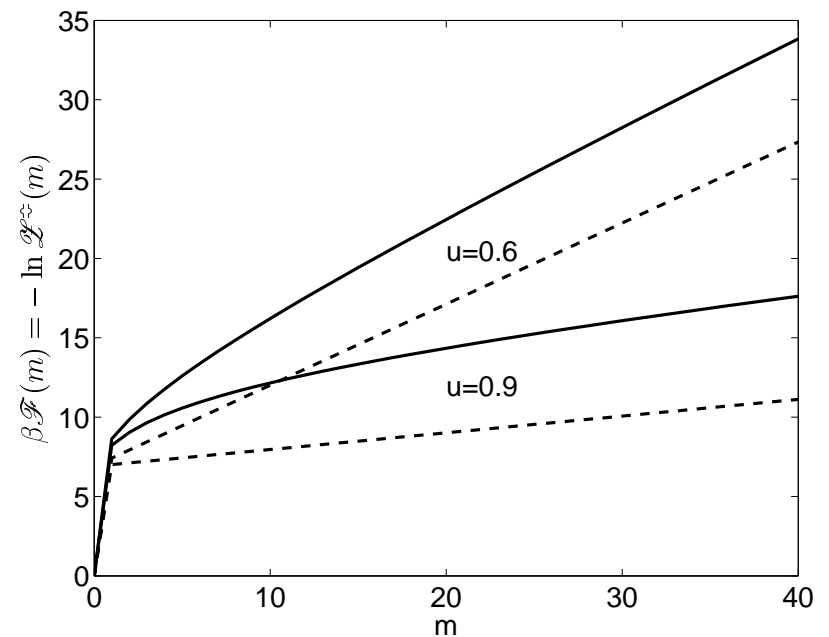
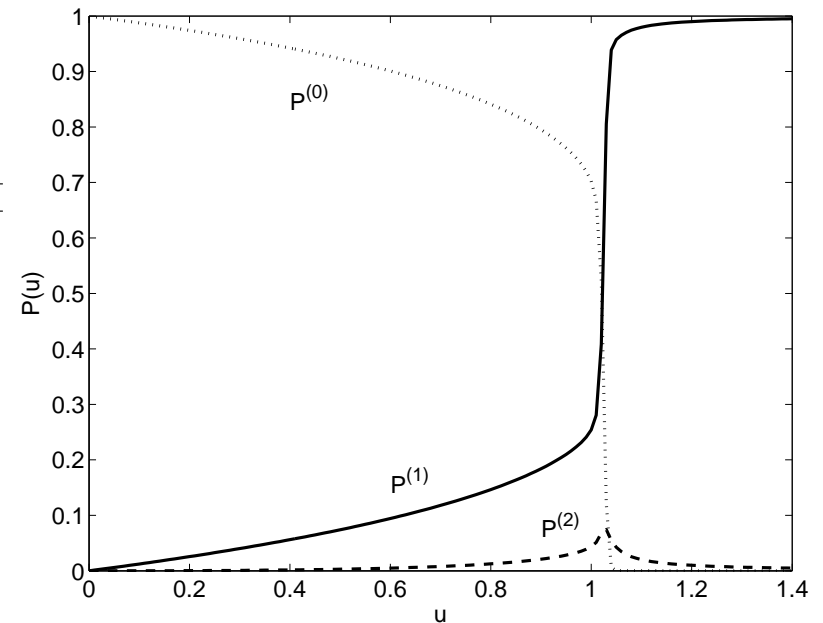
$$\begin{array}{l} \cdot\text{TA}\cdot \\ \cdot\text{AT}\cdot \end{array} \Delta G = 0.1 k_B T \wedge \begin{array}{l} \cdot\text{GC}\cdot \\ \cdot\text{CG}\cdot \end{array} \Delta G = -3.9 k_B T$$

Loop initiation:  $\sigma_0 \simeq 10^{-3 \dots -5} \triangleq 7 \dots 12 k_B T$

Loop closure exponent:  $c \approx 1.76^{(1)}$

Zippering rate:  $k^{-1} \simeq 20 \dots 100 \mu\text{sec}$

Bubble lifetime:  $\tau_{\text{bubble}} \simeq 1 \text{ msec}$



Cooperativity factor  $\sigma_0$  or ring factor  $\epsilon$

Classically,

$$\sigma_0 = \exp\left(-\frac{F_s}{RT}\right) \approx 10^{-5} \dots 10^{-4}, \quad F_s \approx 8\text{kcal/mol}$$

Claim:  $F_s = T\Delta S$ :

$$\exp\left(-\frac{\Delta S_{\text{TA/AT}}}{R}\right) = 1.19 \times 10^{-5}, \quad \text{Heteropolymer, best value: } \sigma_0 = 1.26 \times 10^{-5}$$

$$\exp\left(-\frac{\Delta S_{\text{AT/GC}}}{R}\right) = 3 \times 10^{-6}, \quad \text{GC-rich: } \sigma_0 = 3.5 \times 10^{-6}$$

$$\exp\left(-\frac{\Delta S_{\text{GC/GC}}}{R}\right) = 9.7 \times 10^{-7}, \quad \text{GC homopolymer}$$

Ring factor (nicked DNA!):

$$\sigma_0 = \xi \exp\left(-\frac{\Delta G^{\text{ST}}}{RT}\right)$$

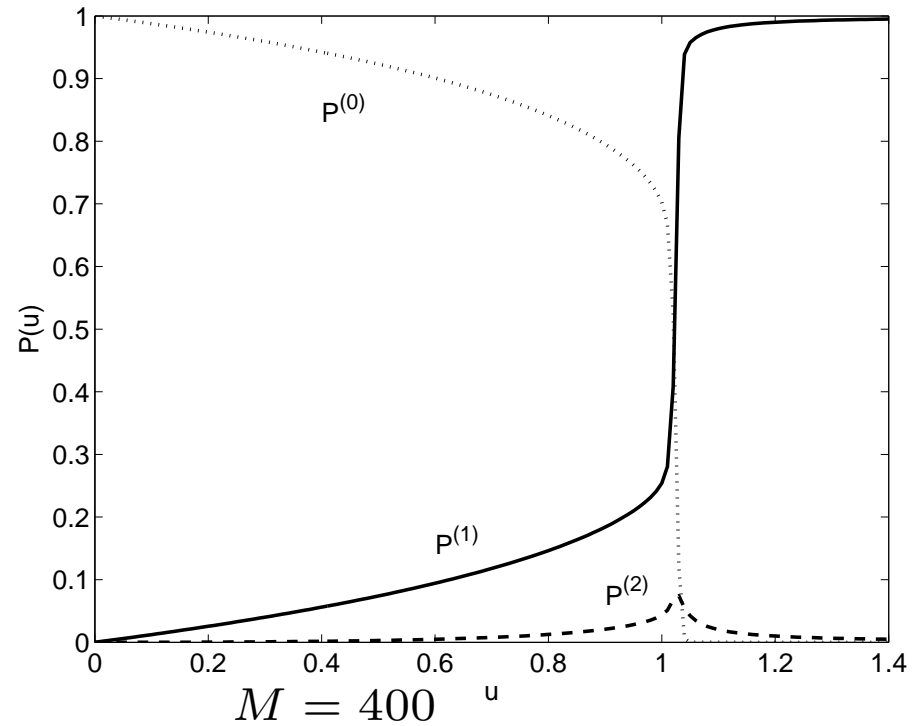
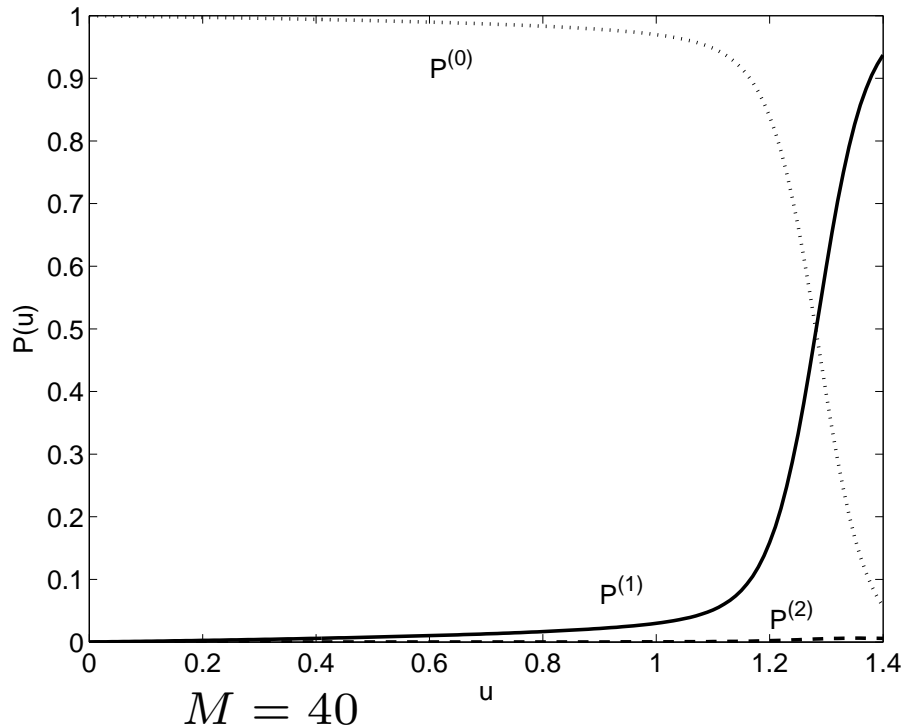
such that for the lowest stacking free energy,  $\sigma_0 \approx 5.6 \times 10^{-4}$

# Poland-Scheraga Free Energy

$$\mathcal{Z}(M) = 1 + \sigma_0 \mathcal{Z}^{(1)}(M) + \sigma_0^2 \mathcal{Z}^{(2)}(M) + \dots$$

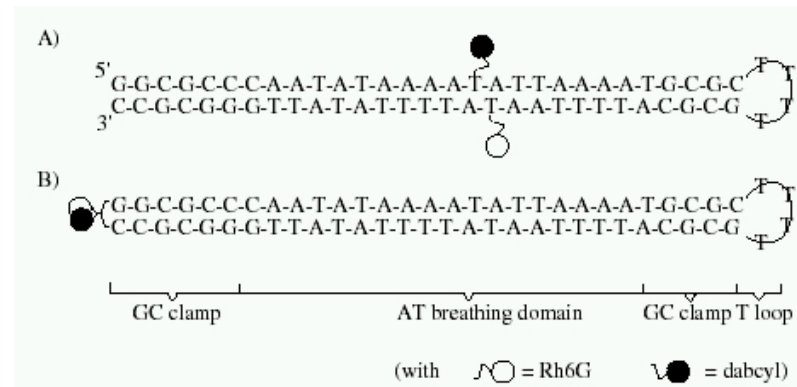
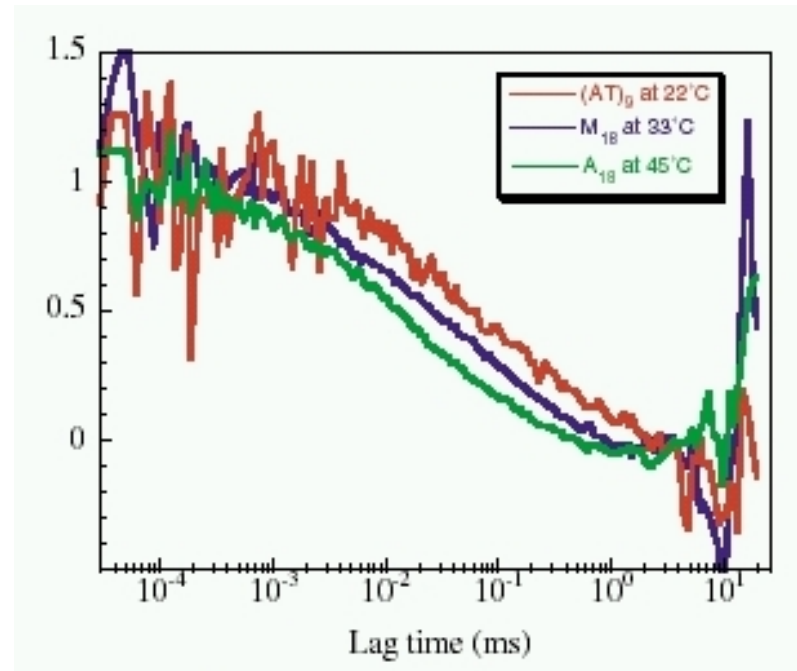
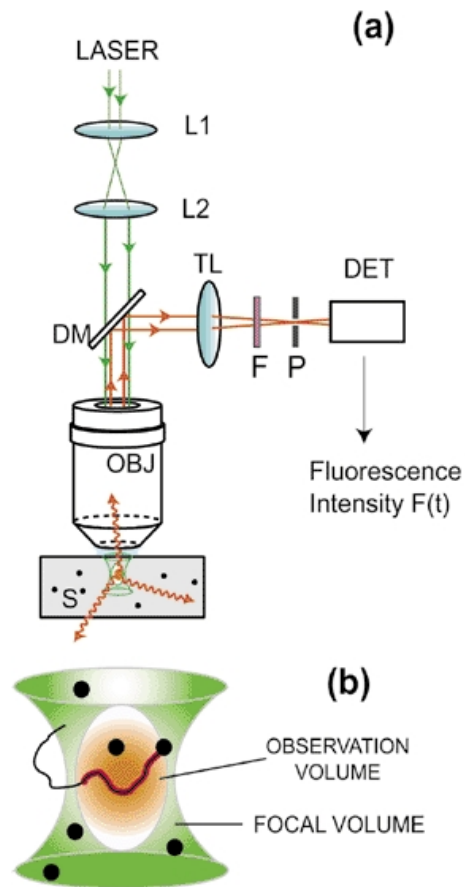
$$\mathcal{Z}^{(1)}(M) = \sum_{m=1}^M (M - m + 1)g(m) \quad \therefore g(m) = u^m (1 + m)^{-c}$$

$$\mathcal{Z}^{(2)}(M) = \frac{1}{2} \sum_{m=1}^{M-1} \sum_{m'=1}^{M-m-1} (M - m - m' + 1)(M - m - m')g(m)g(m')$$





# Watching a single DNA breathe



# Master equation for bubble breathing

$$\frac{\partial}{\partial t} P(m, t) = t^+(m-1)P(m-1, t) + t^-(m+1)P(m+1, t) - (t^+(m) + t^-(m))P(m, t)$$

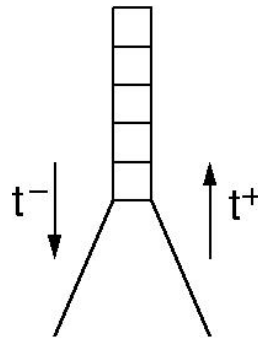
Unzipping rate:

$$t^+(m) = ku \left( \frac{1+m}{2+m} \right)^c$$

$$\therefore t^+(0) = 2^{-c} k \sigma_0 u$$

Zippering rate:

$$t^-(m, n) = k$$

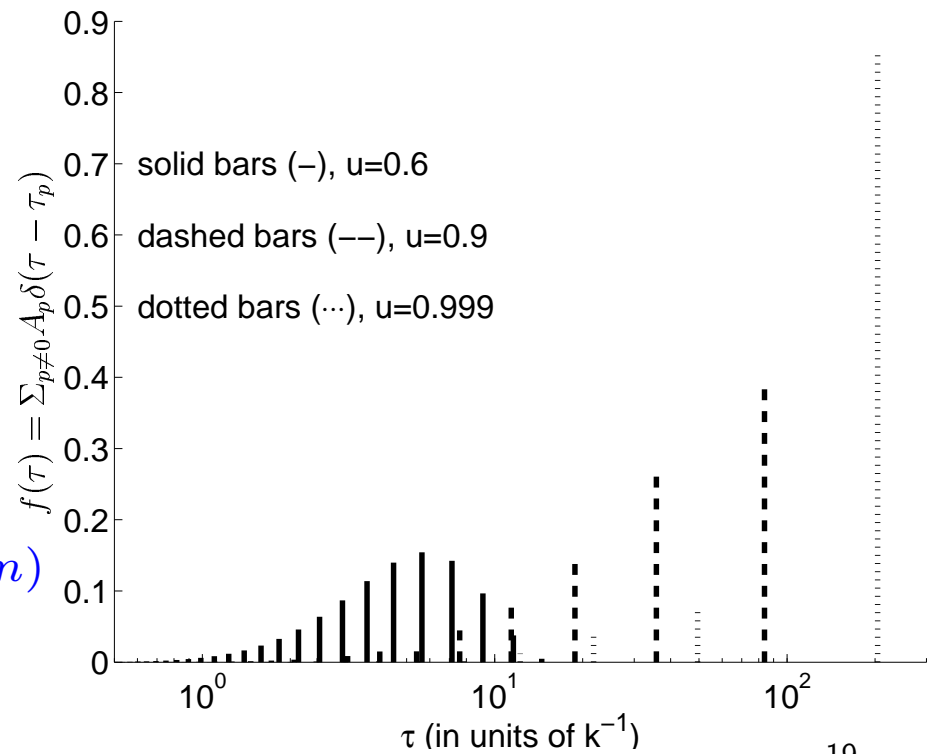


Detailed balance:

$$\mathcal{Z}^\diamond(m) = \sigma_0 u^m (1+m)^{-c}$$

$$t^+(m-1)\mathcal{Z}^\diamond(m-1) = t^-(m)\mathcal{Z}^\diamond(m)$$

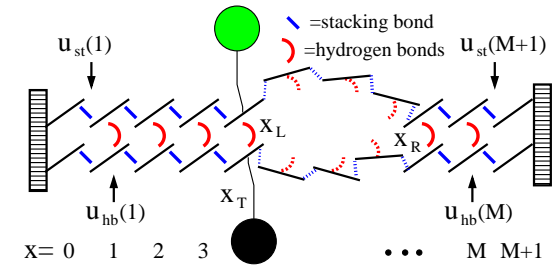
$$P_{\text{eq}}(m)/P_{\text{eq}}(m') = \mathcal{Z}^\diamond(m)/\mathcal{Z}^\diamond(m')$$



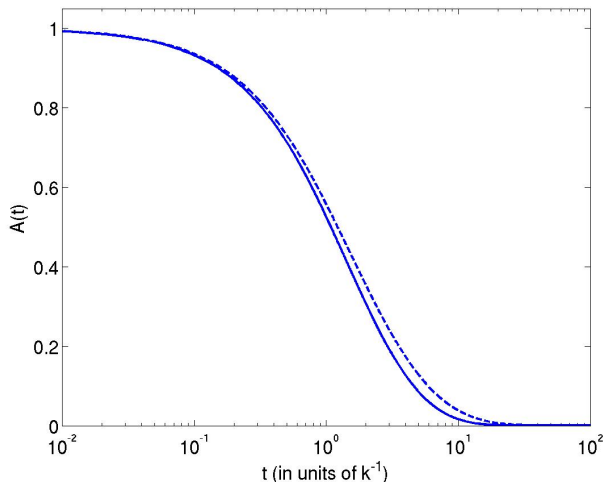
# Experiment measures a tagged base-pair:

Random variable  $I(t) \in \{0, 1\}$

(Similar if  $x' \pm \Delta$  broken bps necessary for signal)

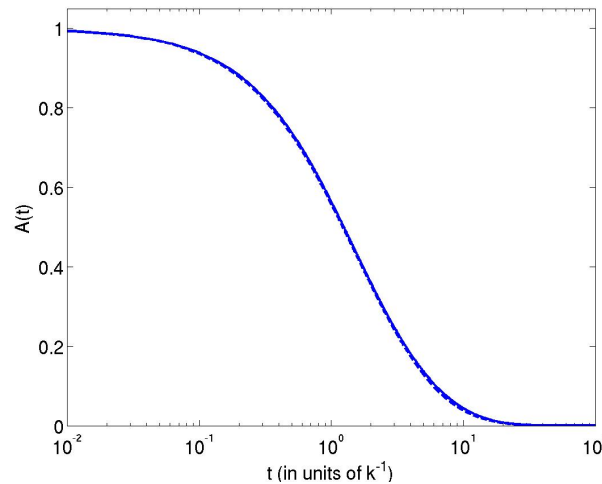


Size  $\langle m(t)m(0) \rangle - \langle m \rangle^2$  (---) vs blinking autocorrelation  $\langle I(t)I(0) \rangle - \langle I \rangle^2$  (—):



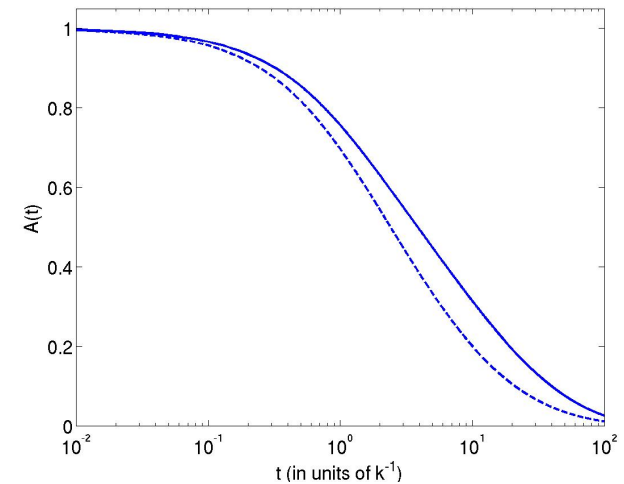
Tag close to clamp

$$u = 0.6$$



Centre-tag

$$u = 0.6$$

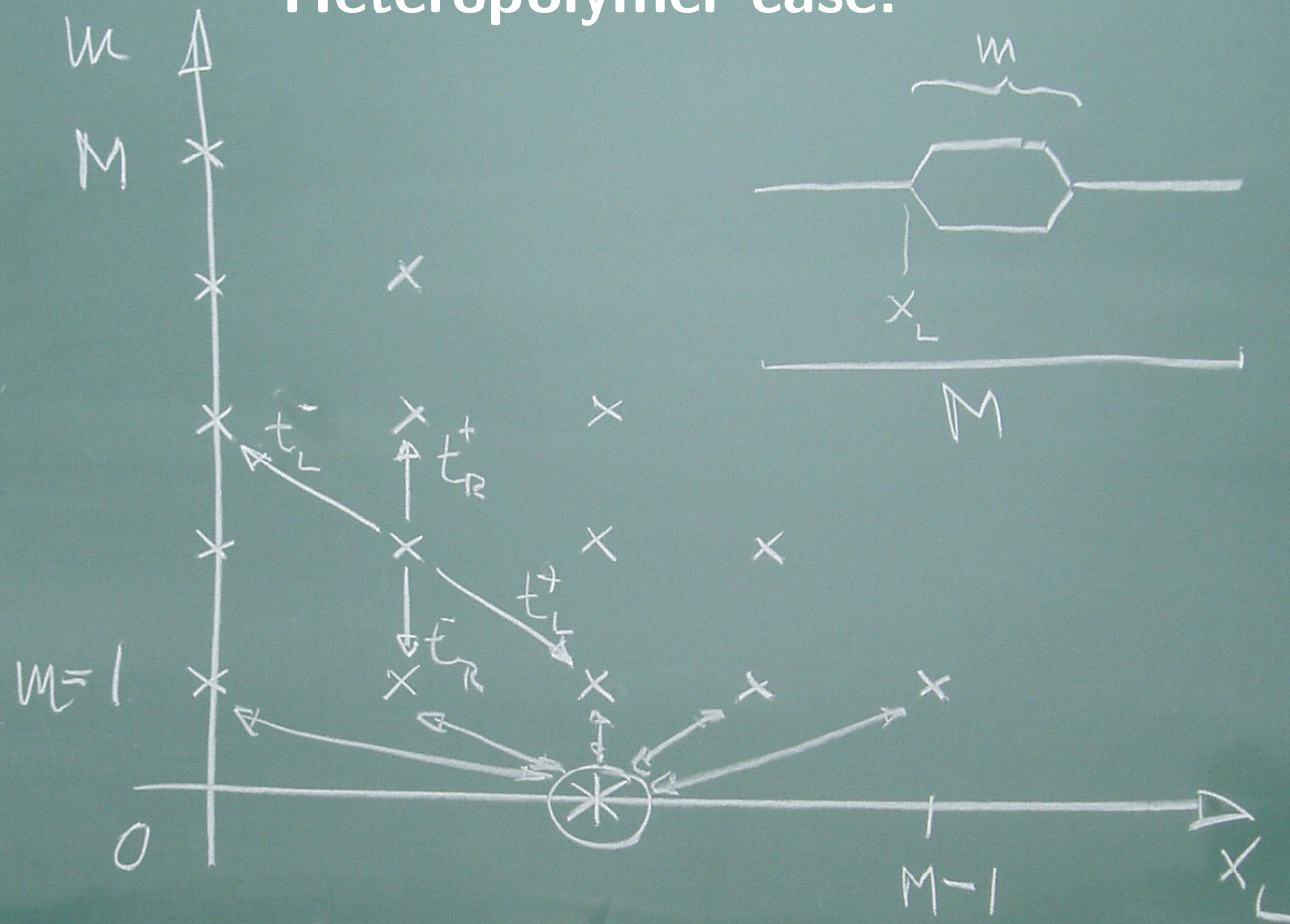


Centre-tag

$$u = 0.9$$



# Heteropolymer case:

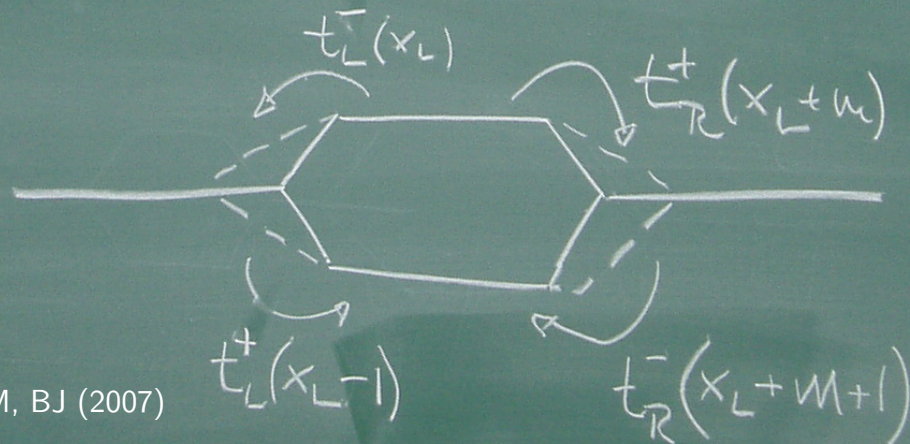


Sequence dependence:

$$u, \sigma_0 \rightarrow u(x), \sigma_0(x)$$

$$t^\pm \rightarrow t_{R/L}^\pm(x_{R/L})$$

$$P \rightarrow P(m, x_L, t)$$



## Partition sum for heteropolymeric DNA

$$\mathcal{Z}(x_L, m) = \frac{2^c \xi}{(1+m)^c} \prod_{x=x_L+1}^{x_L+m} u_{\text{hb}}(x) \prod_{x=x_L+1}^{x_L+m+1} u_{\text{st}}(x)$$

Where the weights correspond to the free energy of base pairing (2x) and stacking (10x):

$$u_{\text{hb}}(x) = \exp\{-\epsilon_{\text{hb}}(x)/[k_B T]\}, \quad u_{\text{st}}(x) = \exp\{-\epsilon_{\text{st}}(x)/[k_B T]\}$$

Values @ 37 °C and 100 mM NaCl:

$$\epsilon_{\text{hb}}(\text{AT}) = 1.0k_B T, \quad \epsilon_{\text{hb}}(\text{GC}) = 0.2k_B T$$

$$\epsilon_{\text{st}}(\text{TA/AT}) = -0.9k_B T, \quad \epsilon_{\text{st}}(\text{GC/CG}) = -4.1k_B T$$

Note that @  $T = 37^\circ\text{C}$ ,  $k_B T = 0.62\text{kcal/mol}$

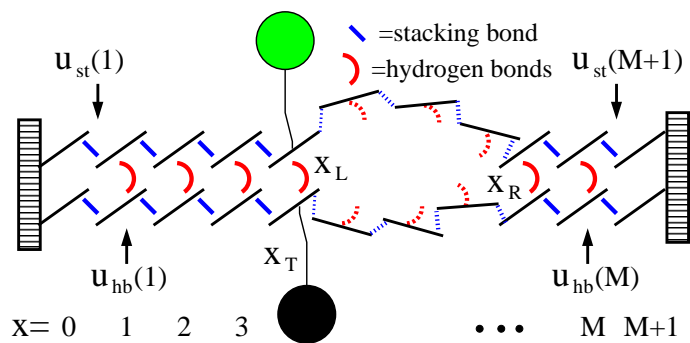
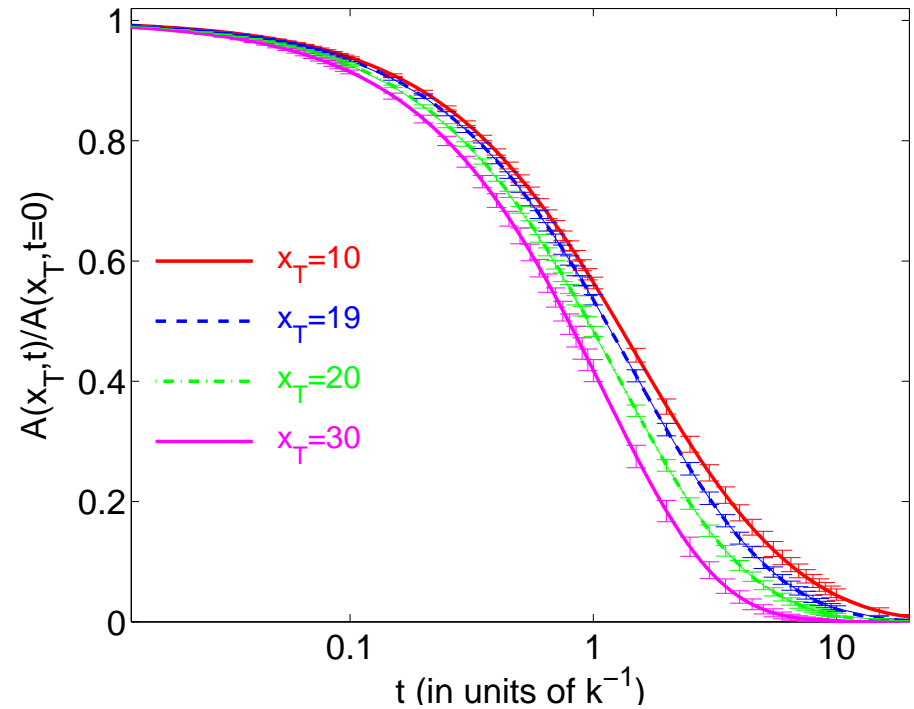
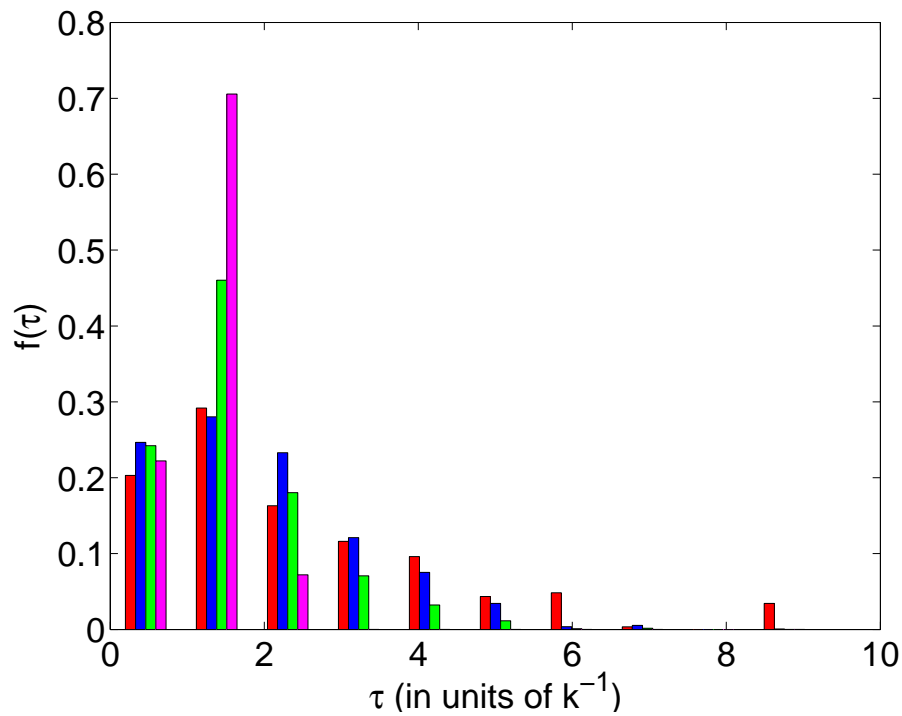
Table 1. Stacking  $\Delta G_{KL}^{ST}$  and base-pairing  $\Delta G^{BP}$  parameters used in our calculations\*

$\Delta G_{KL}^{ST}$	5' KL	A	T	G	C
	A	-1.50	-1.73	-1.45	-2.20
5'	T	-0.58	-1.50	-0.94	-1.82
	G	-1.82	-2.20	-1.83	-2.56
	C	-0.94	-1.45	-1.30	-1.83
$\Delta G^{BP}$	A·T		0.65		
	G·C		0.13		

\* Stacking and base-pairing parameters in kcal/mol correspond to 37°C, 0.1 M NaCl.

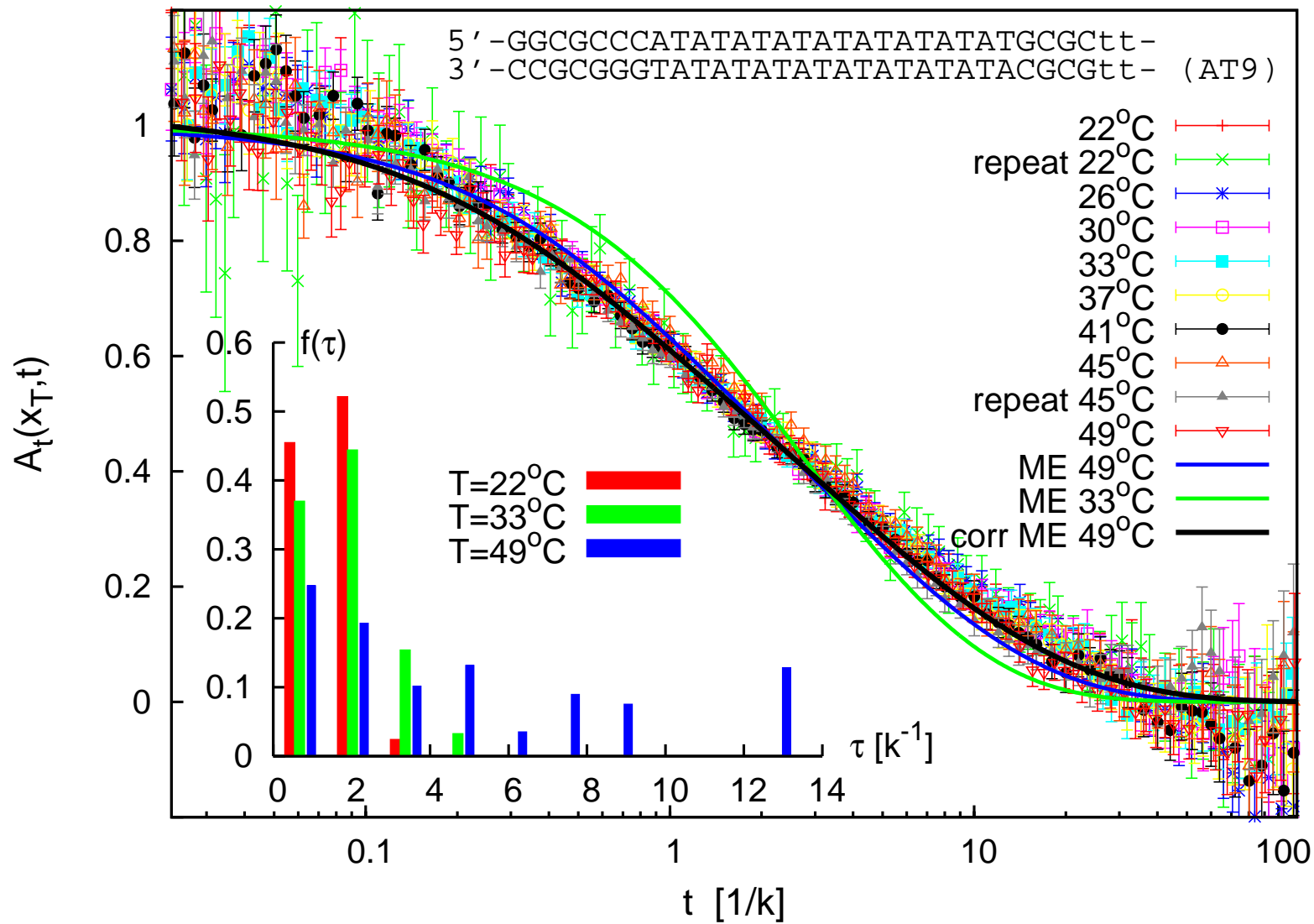


# Breathing dynamics is sensitive to sequence



AAAAAAAAAA AAAAAAAAAA AGGGGGGGGGG CGGGGGGGG  
 TTTTTTTTTT TTTTTTTTTT TCCCCCCCCC CCCCCCCC  
 0123456789012345678901234567890123456789

# Fluorescence autocorrelation function

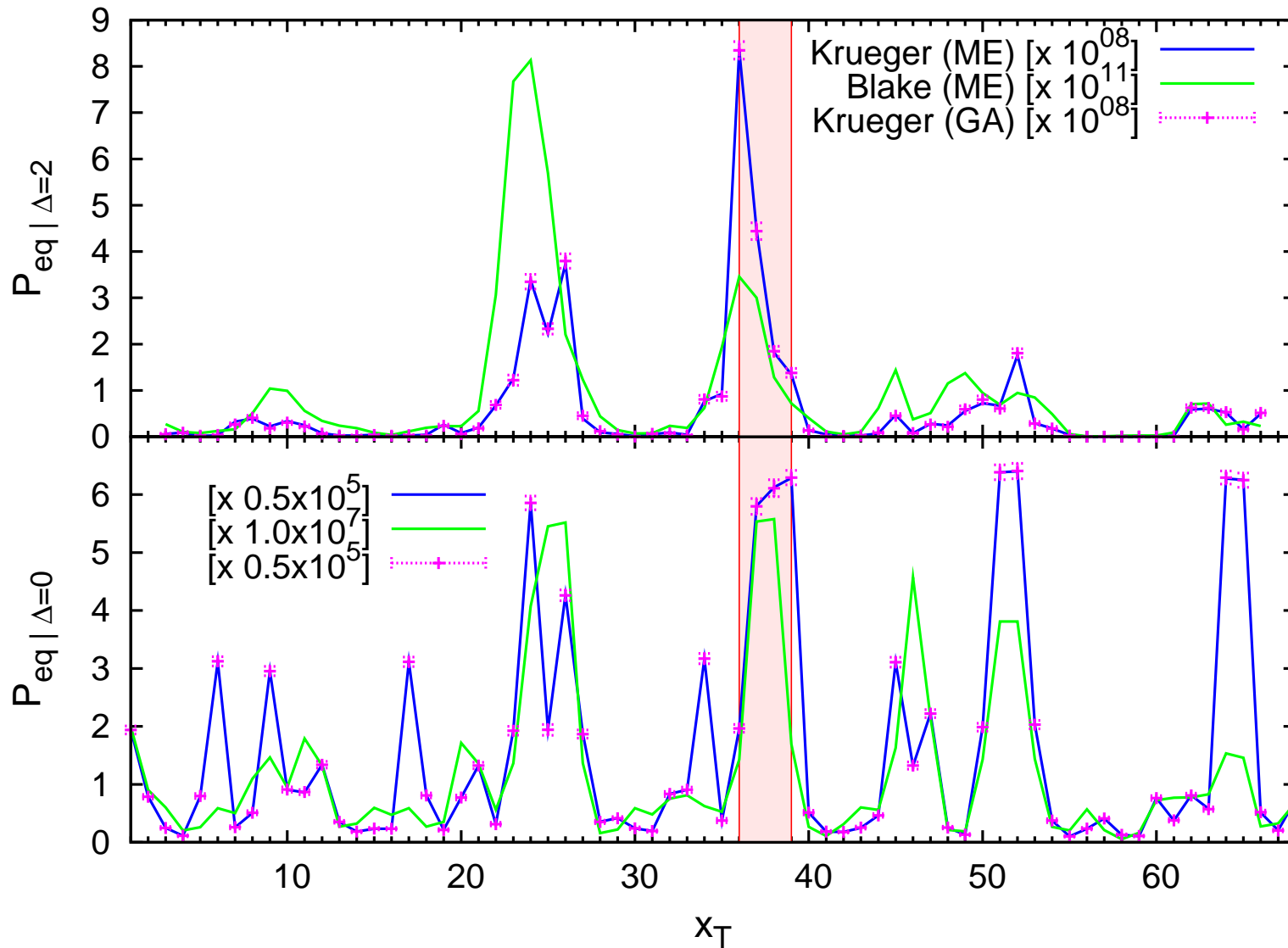
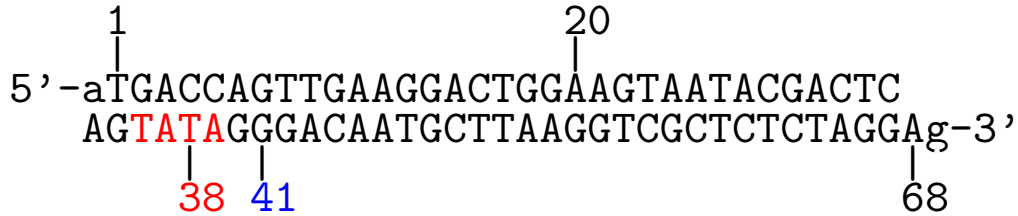


# Phage T7 promoter sequence

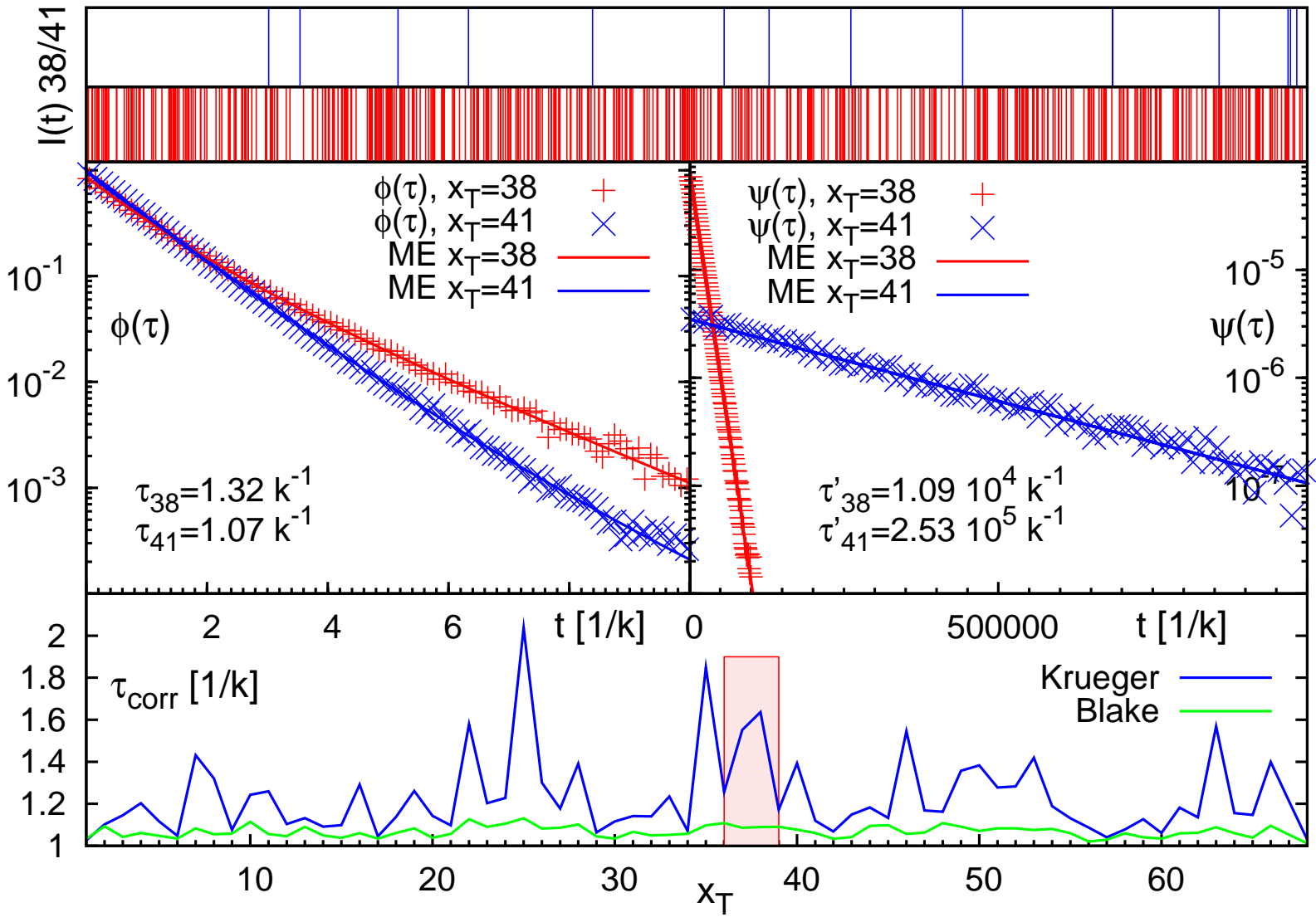
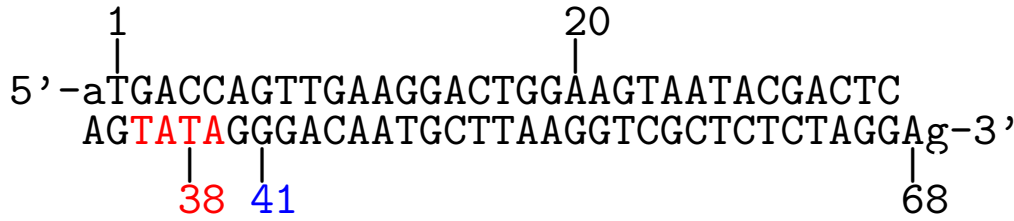
```
      1                20
      |                |
5'-aTGACCAGTTGAAGGACTGGAAGTAATACGACTC
  AGTATAGGGACAATGCTTAAGGTCGCTCTCTAGGAg-3'
      | |                |
      38 41                68
```



# Bubble opening probability

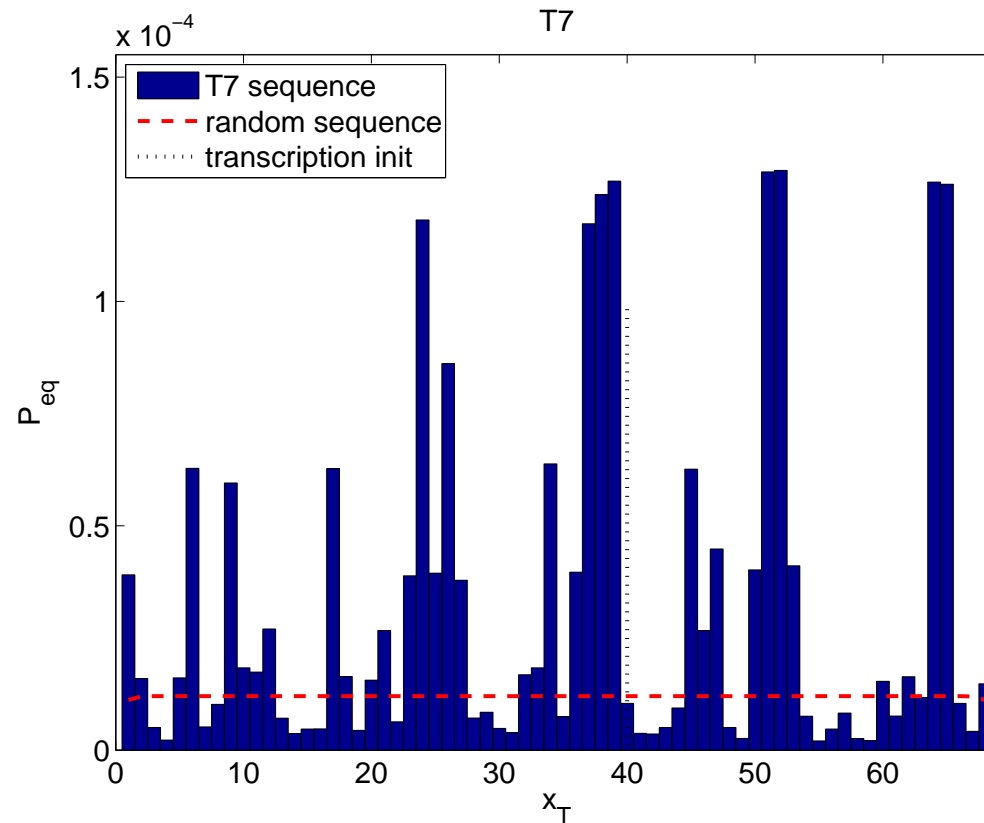
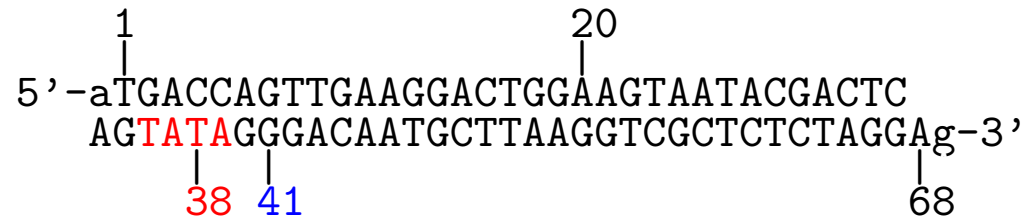


# Fluorescence repeat & time



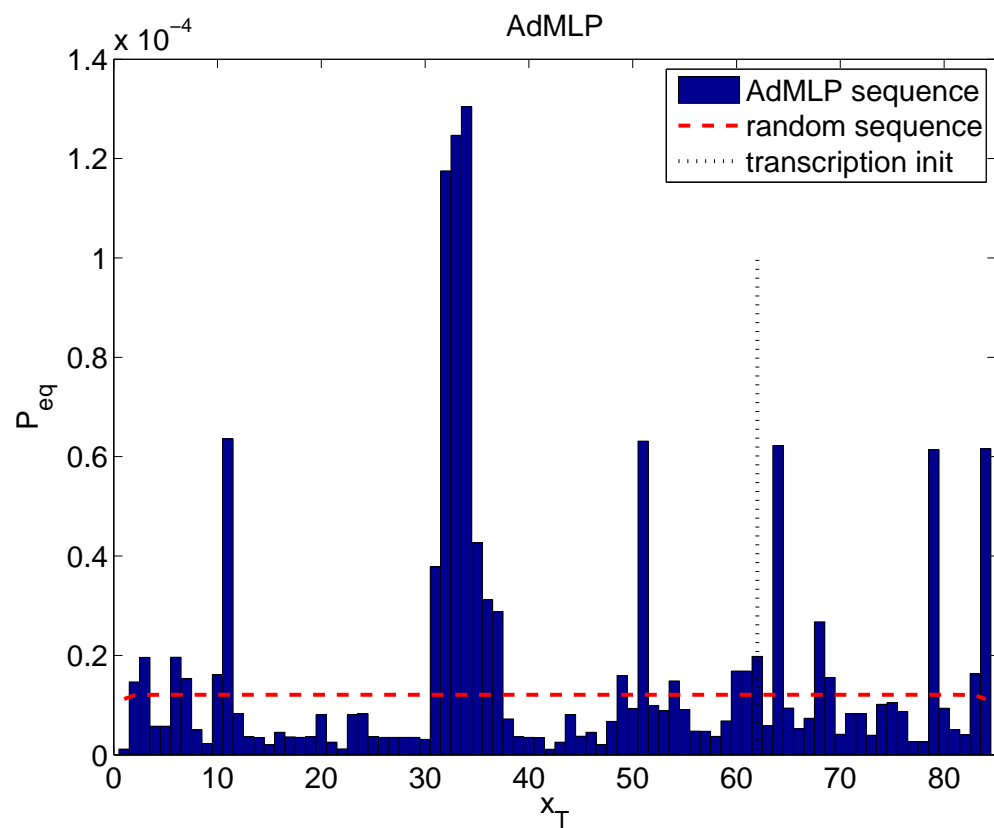
# Biological relevance of DNA breathing

Bacteriophage T7 core promoter:

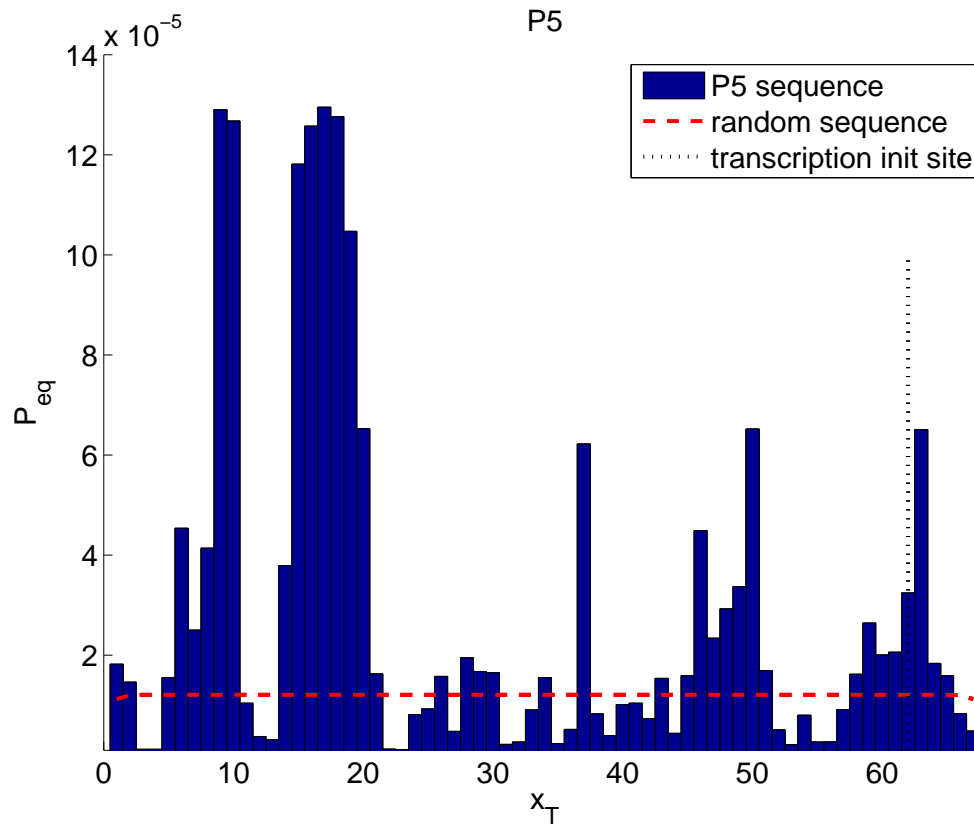




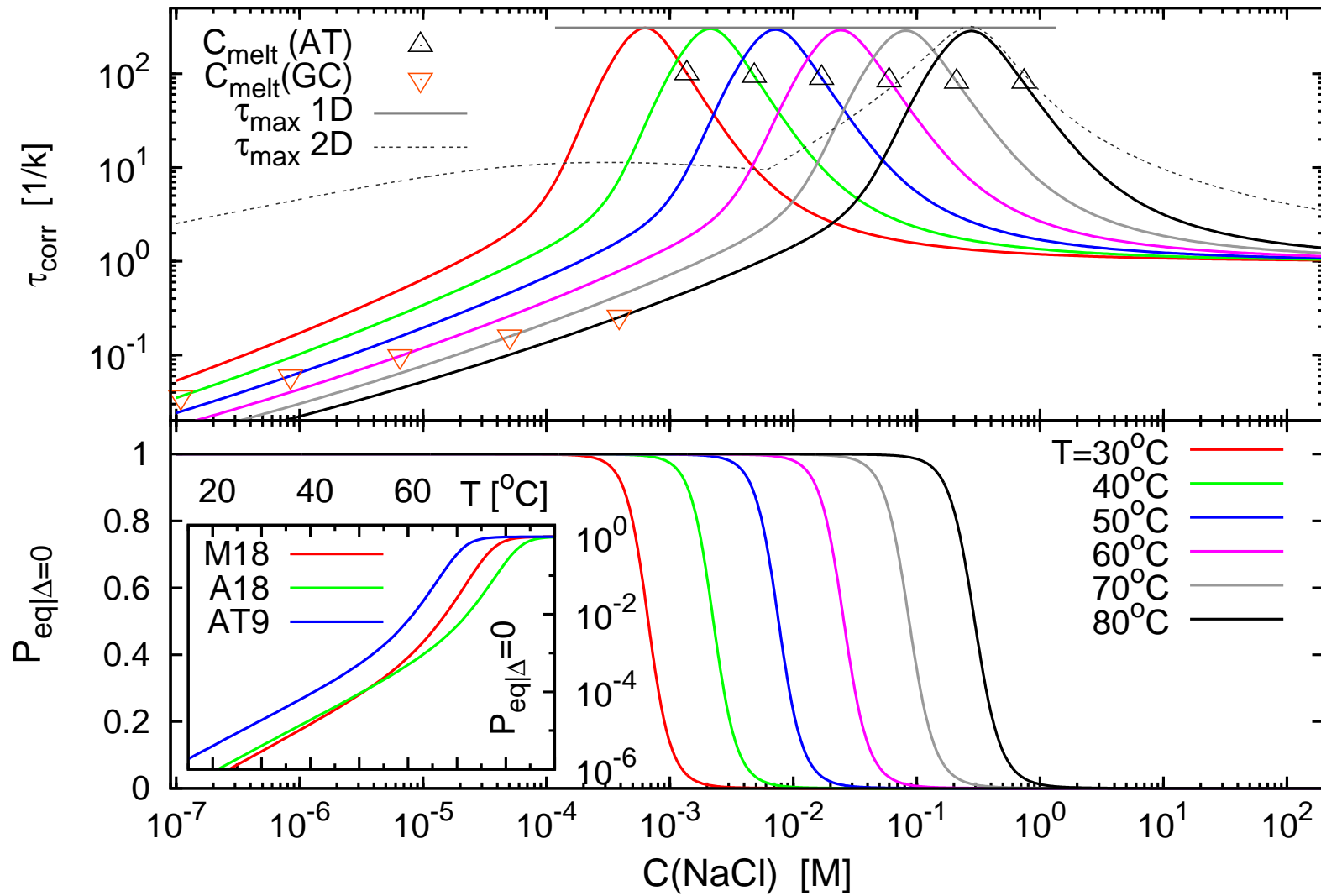
# Adenovirus Major Late Promoter:



Adeno Associated Viral P5 promoter ([YinYang 1](#) motif):

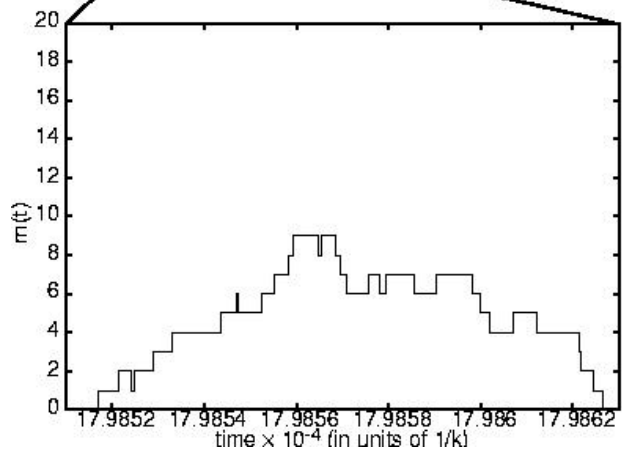
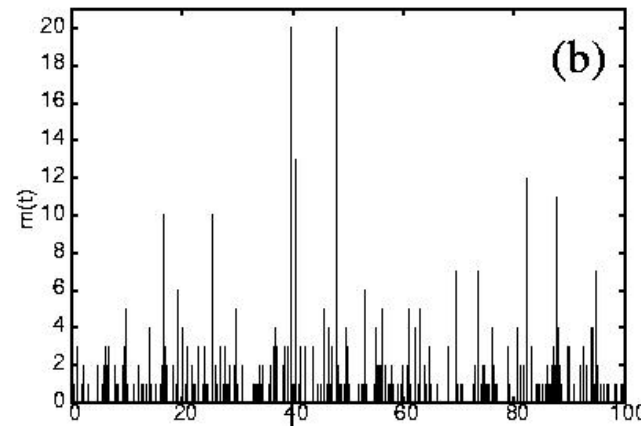
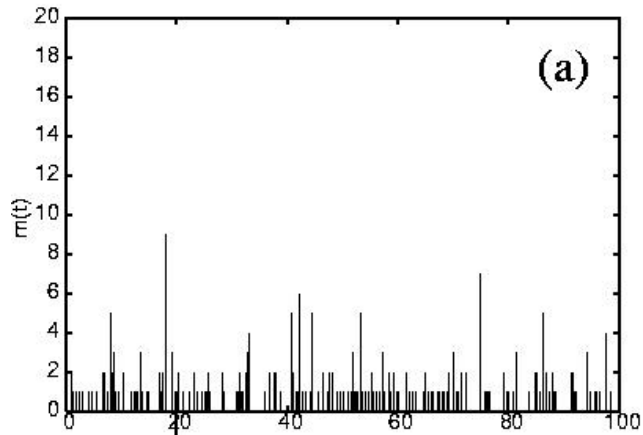


# Salt dependence of correlation time (AT9)

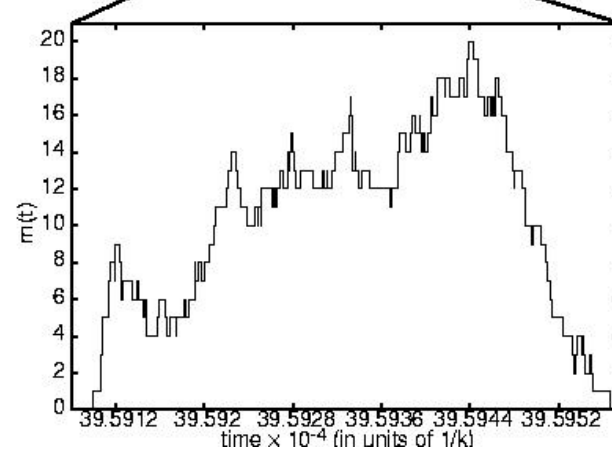


# Single bubble time series—Stochastic Gillespie algorithm

Reaction PDF  $P(\tau, \mu; m) = t^\mu(m) \exp \{ - (t^+(m) + t^-(m)) \tau \} \therefore \mu \in \{+, -\}$



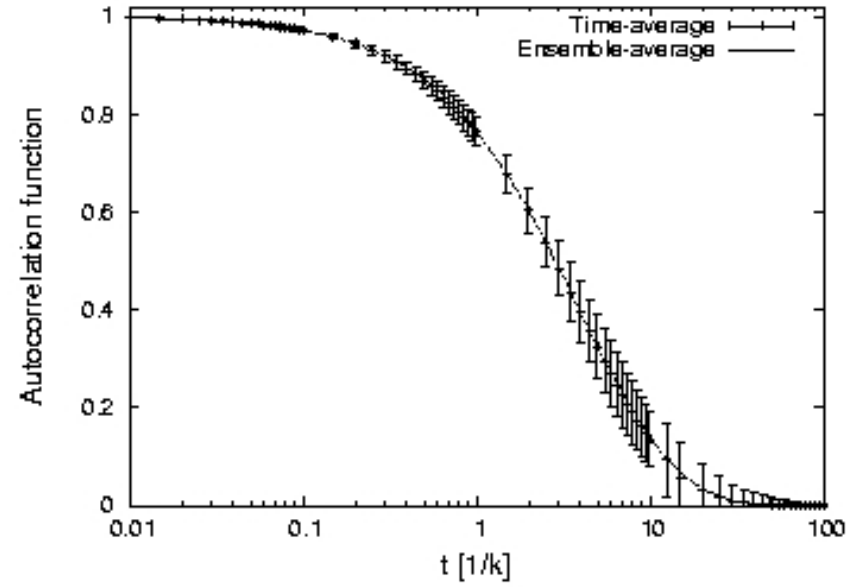
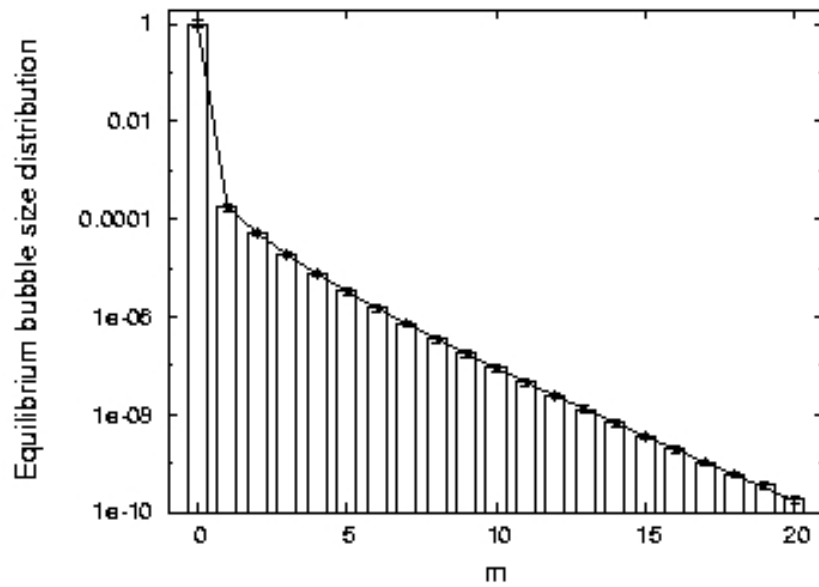
$u = 0.6$



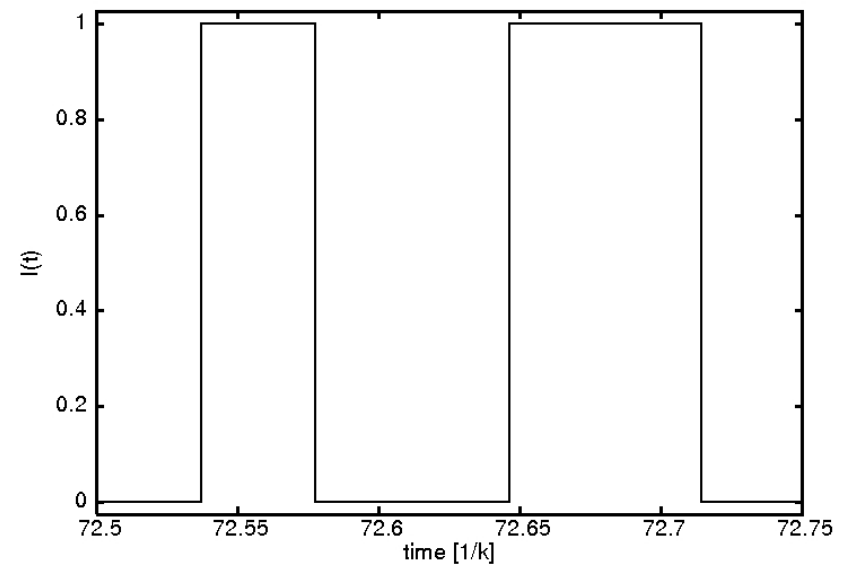
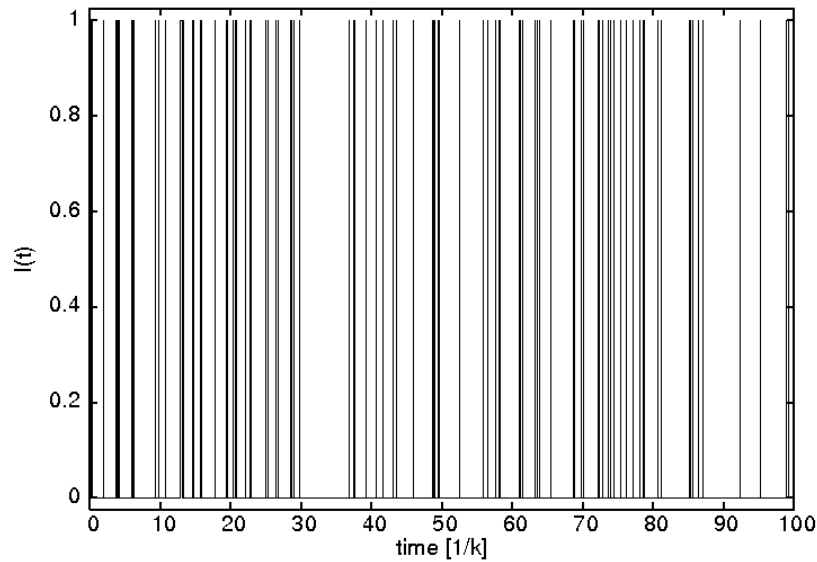
$u = 0.9$



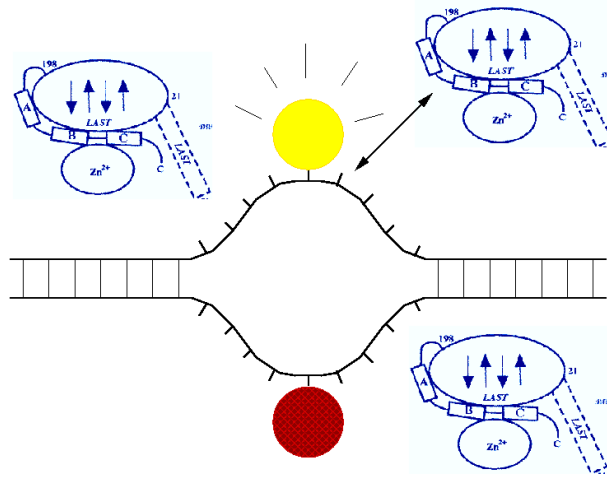
# Histogram of equilibrium bubble sizes and autocorrelation:



# Heteropolymer domain with single tag:



# DNA and single-stranded DNA binding proteins (SSBs):



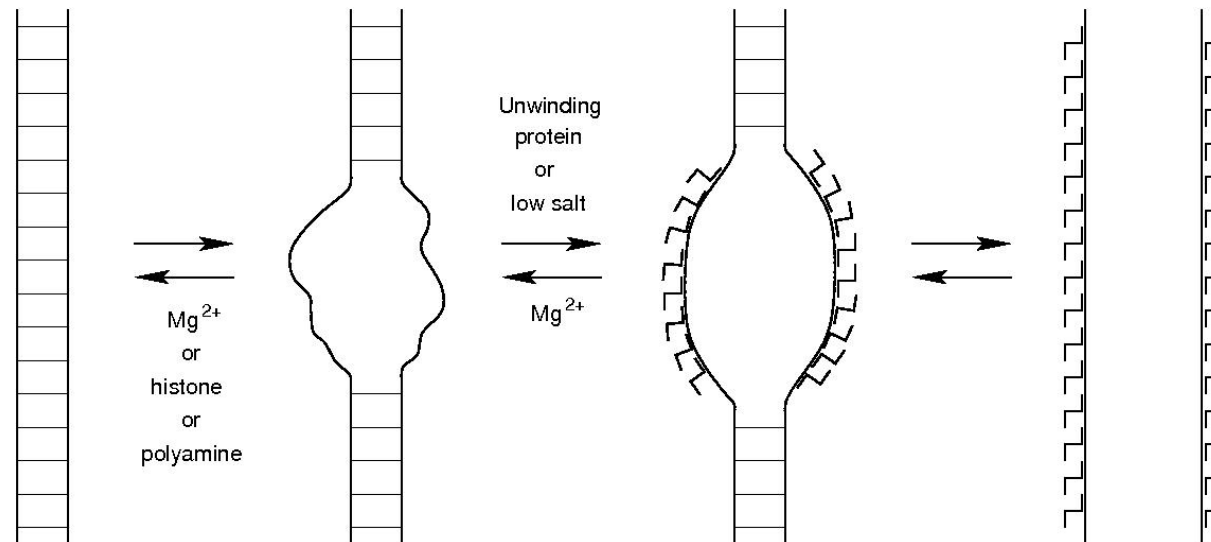
Binding strength  $\kappa = c_0 K^{\text{eq}}$

Equilibrium constant  $K^{\text{eq}}$

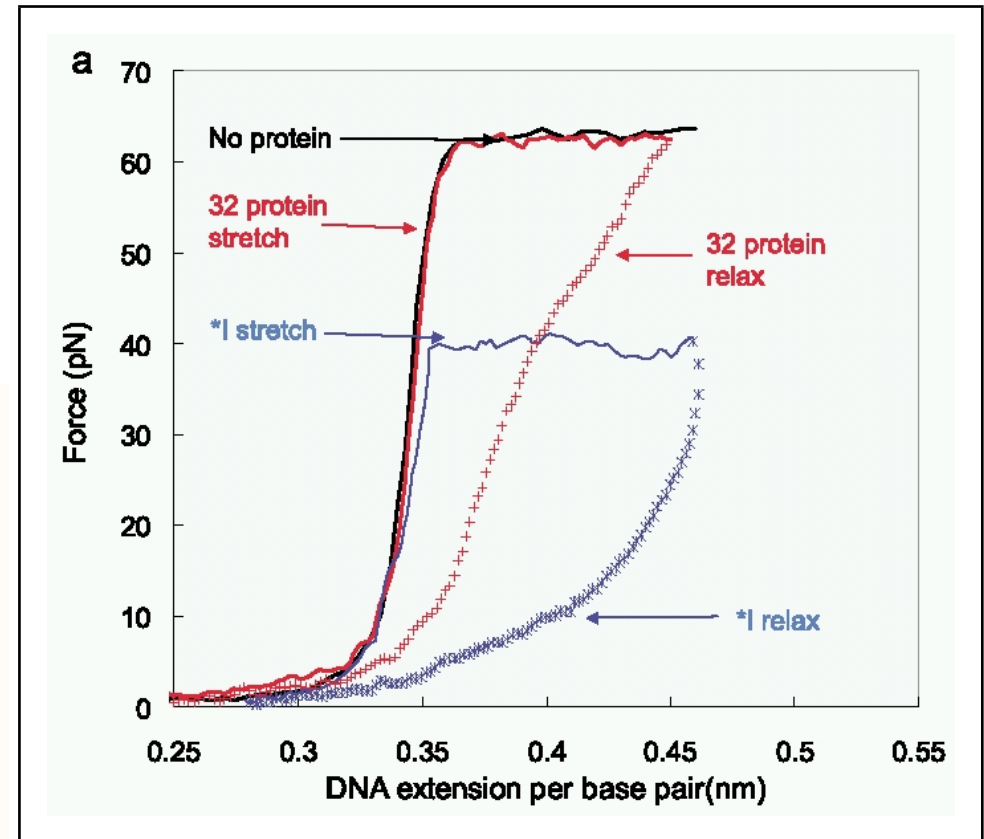
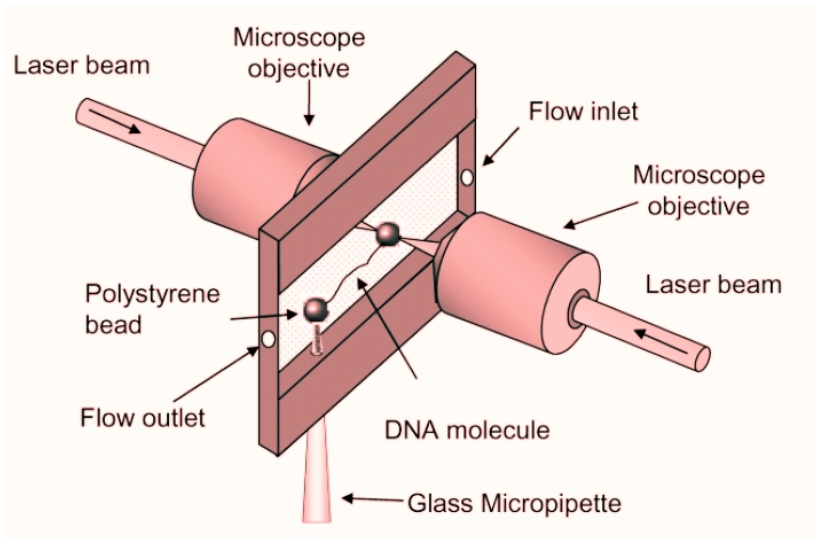
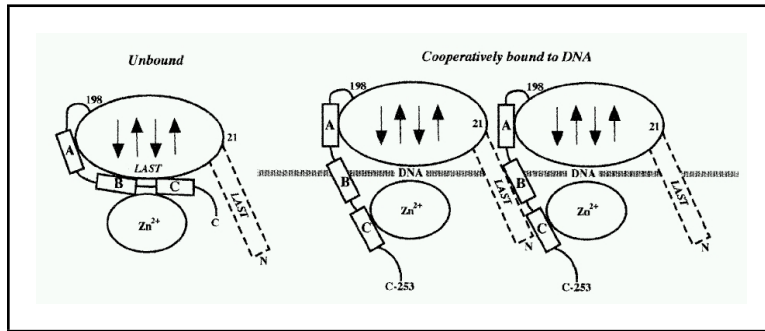
SSB-concentration  $c_0$

SSB-size  $\lambda$  in units of bp

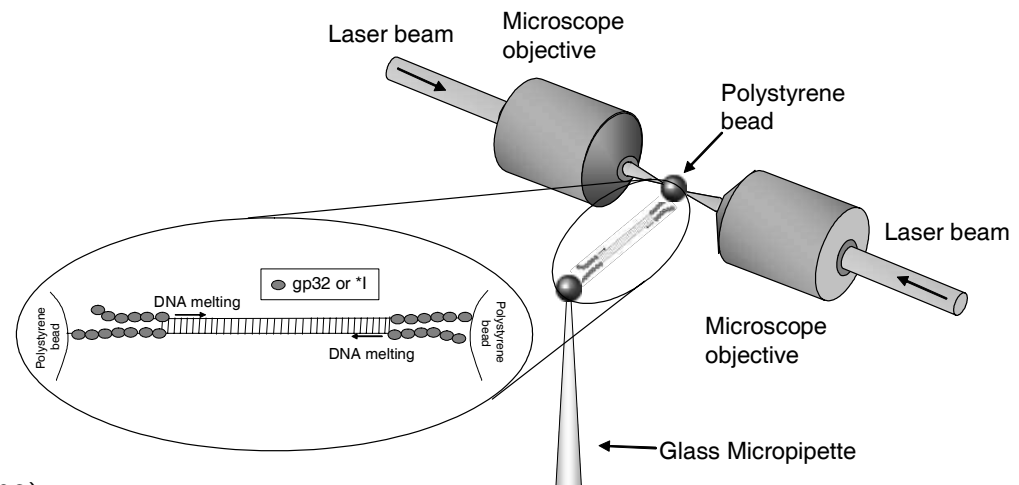
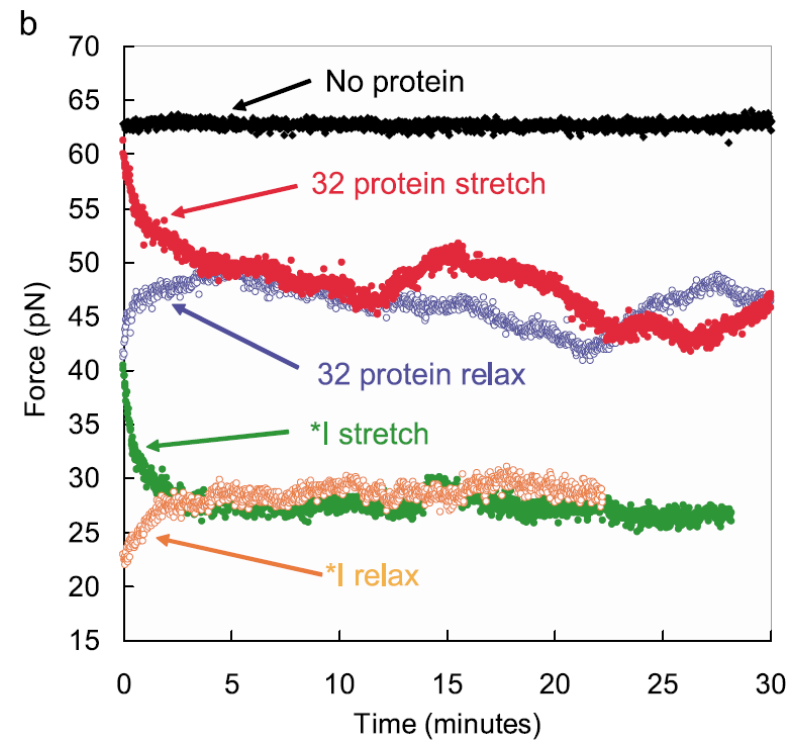
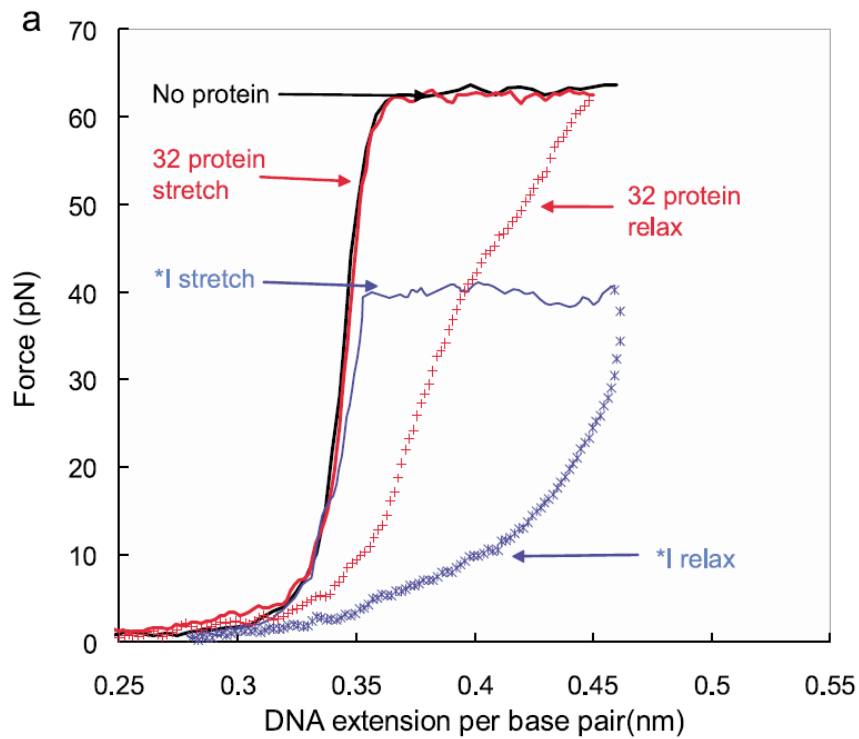
Classical view  
SSB-induced  
denaturation:



# DNA-overstretching experiments in the presence of bacteriophage T4 g32p SSBs

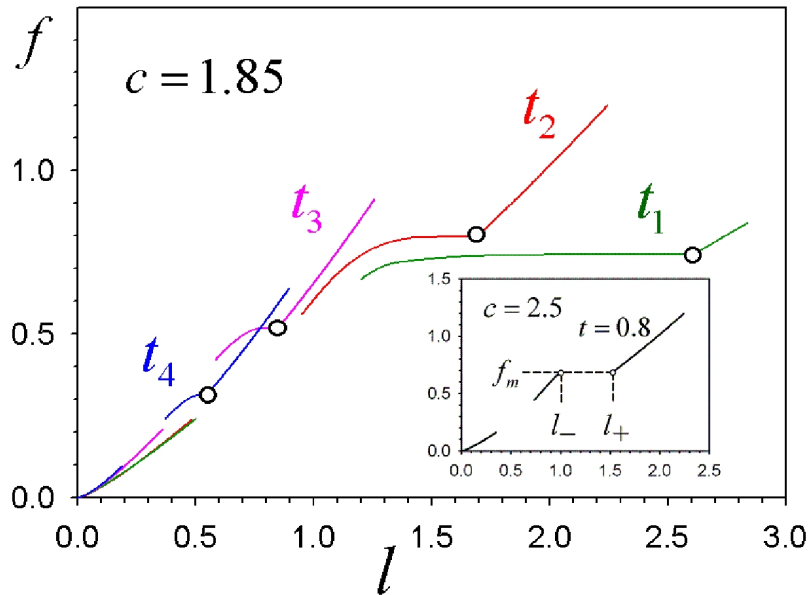
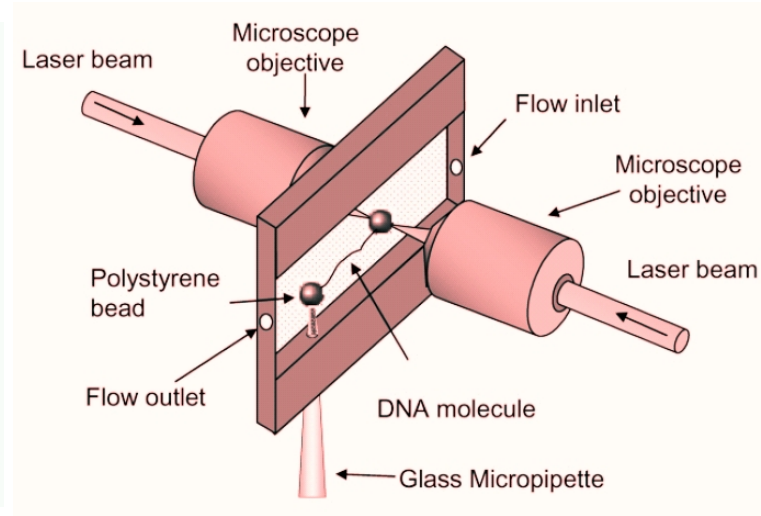
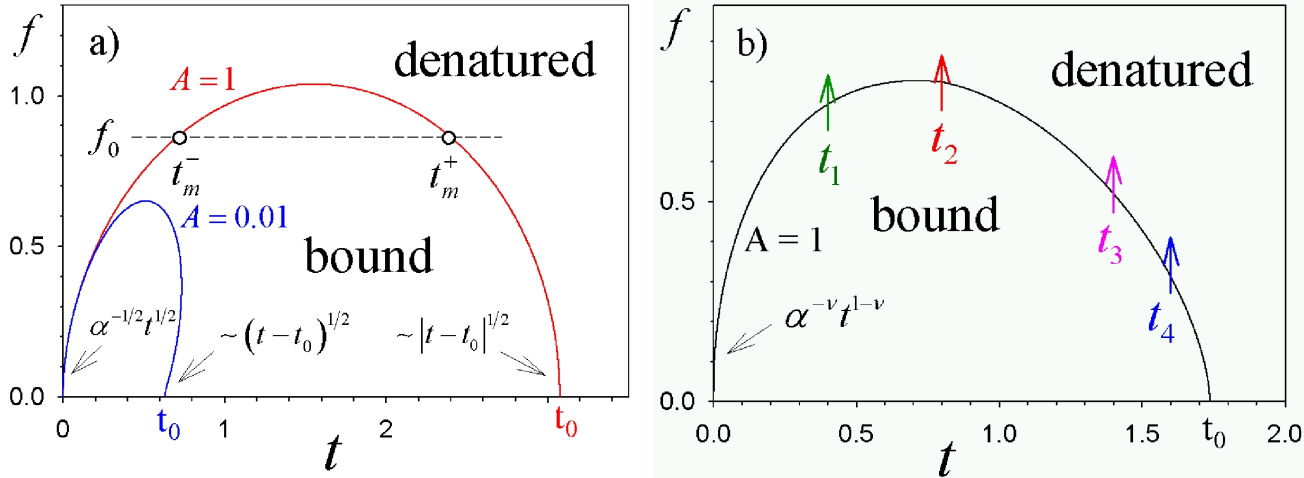


# DNA and single-stranded DNA binding proteins (SSBs):





# Force-induced melting of DNA



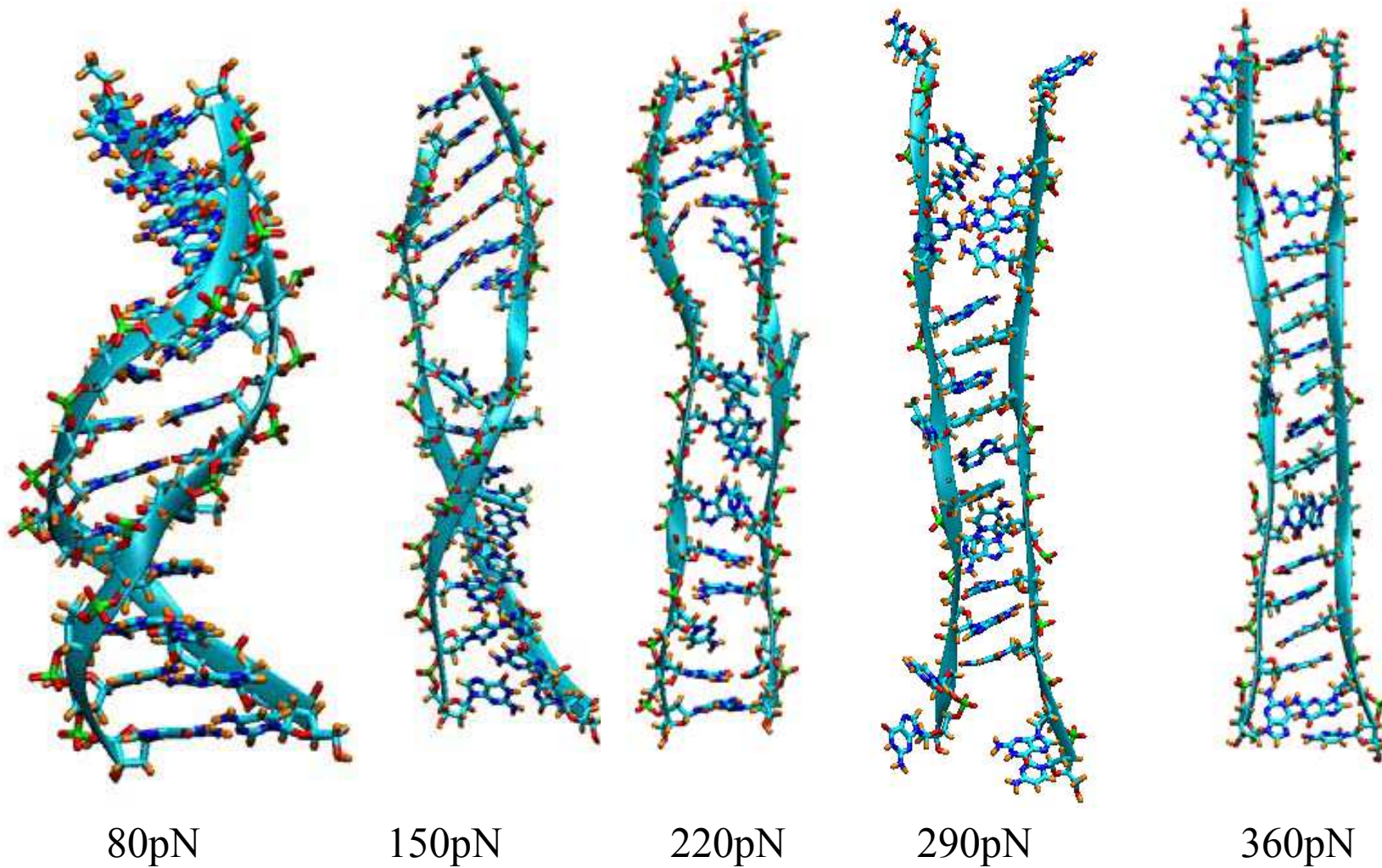
@  $F > 0$  new critical exponent:

$$c = 4\nu - 1/2 \approx 1.85$$

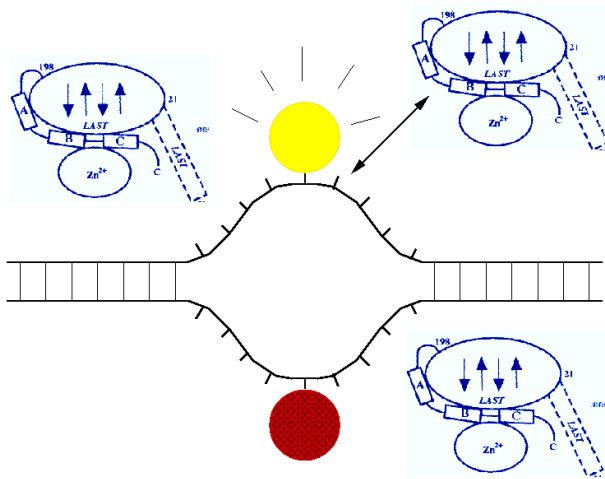
$$(c = 3\nu \approx 1.76 \text{ @ } F = 0)$$

Melting temperature:  $T_m = T_m(F)$

# S-DNA debate



## Breathing bubbles interacting with single-strand binders (SSBs):



Binding strength  $\kappa = c_0 K^{\text{eq}}$

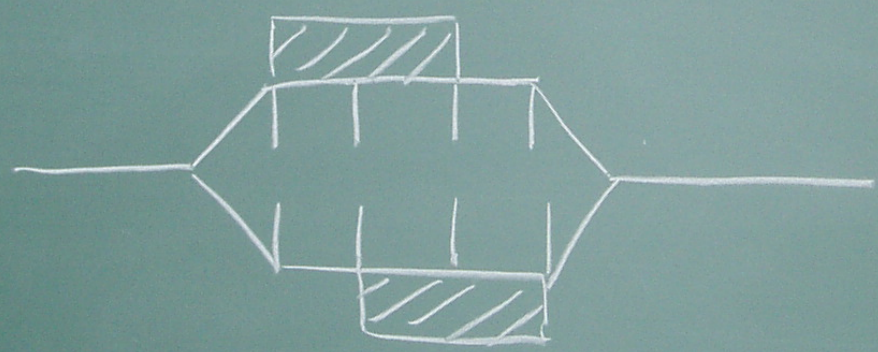
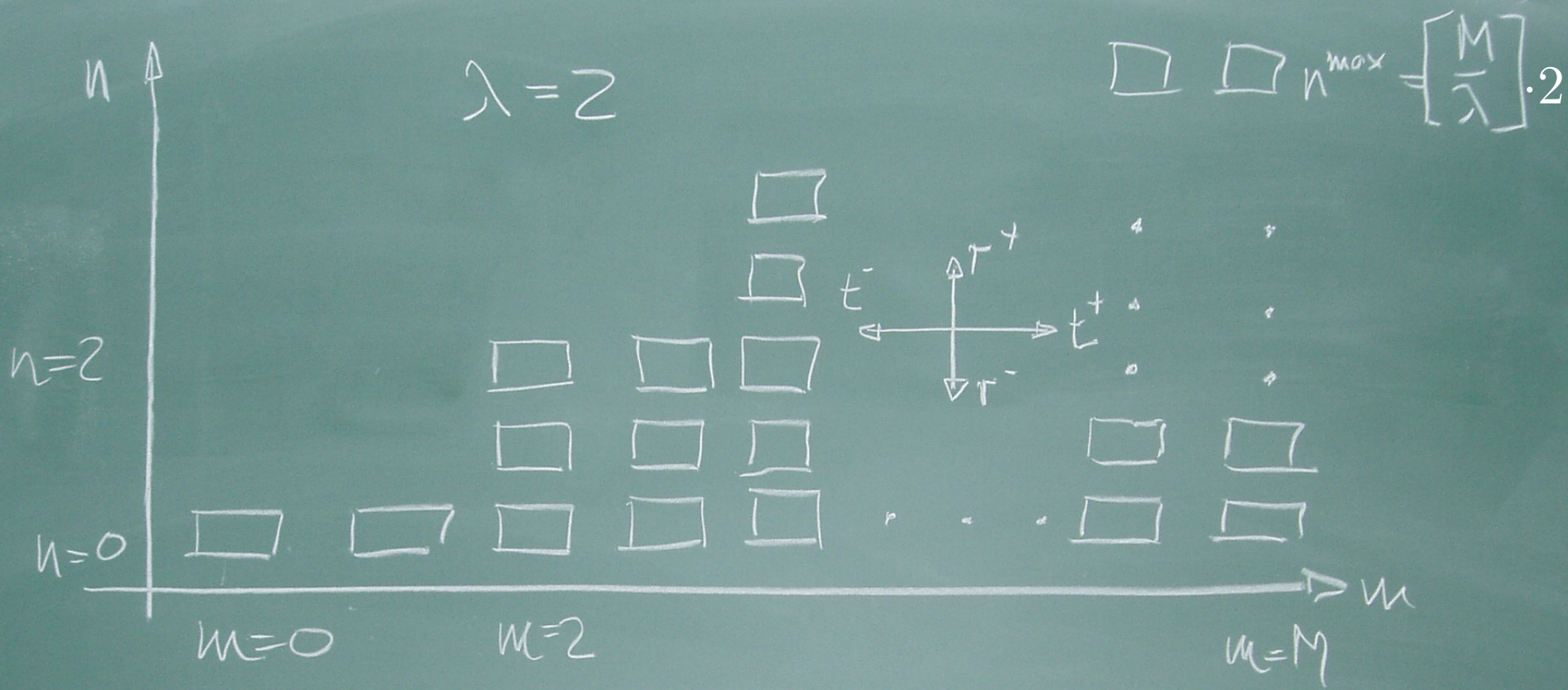
Equilibrium constant  $K^{\text{eq}}$

SSB-concentration  $c_0$

SSB-size  $\lambda$  in units of bp

$$\begin{aligned} \frac{\partial}{\partial t} P(m, n, t) = & \mathbf{t}^+(m-1, n)P(m-1, n, t) + \mathbf{t}^-(m+1, n)P(m+1, n, t) \\ & - \left( \mathbf{t}^+(m, n) + \mathbf{t}^-(m, n) \right) P(m, n, t) \\ & + \mathbf{r}^+(m, n-1)P(m, n-1, t) + \mathbf{r}^-(m, n+1)P(m, n+1, t) \\ & - \left( \mathbf{r}^+(m, n) + \mathbf{r}^-(m, n) \right) P(m, n, t) \end{aligned}$$





Transfer rates fulfil detailed balance:

$$t^+(m-1, n) \mathcal{Z}(m-1, n) = t^-(m, n) \mathcal{Z}(m, n)$$

$$r^+(m, n-1) \mathcal{Z}(m, n-1) = r^-(m, n) \mathcal{Z}(m, n)$$

Partition function:

$$\mathcal{Z}(m, n) = \mathcal{Z}^{\text{bubble}}(m) \mathcal{Z}^{\text{bind}}(m, n)$$

$$\mathcal{Z}^{\text{bubble}}(m) = \sigma_0 u^m (1+m)^{-c} \quad \therefore \quad \mathcal{Z}^{\text{bubble}}(0) = 1, \quad u = e^{-\beta(E - \theta_0 \tau)}$$

$$\mathcal{Z}^{\text{bind}}(m, n) = \Omega^{\text{bind}}(m, n) \kappa^n \quad \therefore \quad \kappa = c_0 v_0 e^{\beta |E_{\text{bind}}|}$$

$$\Omega^{\text{bind}}(m, n) = \sum_{n'=0}^n \omega^{\text{bind}}(m, n') \omega^{\text{bind}}(m, n - n') \left| \begin{array}{l} n - n' \leq n^{\text{max}}/2 \\ n' \leq n^{\text{max}}/2 \end{array} \right.$$

$$\omega^{\text{bind}}(m, n) = \binom{m - (\lambda - 1)n}{n}$$



## Bubble breathing-SSB binding transfer rates

$$t^+(m, n) = t^+(m) = ku \left( \frac{1+m}{2+m} \right)^c \quad \therefore t^+(0) = 2^{-c} k \sigma_0 u$$

$$t^-(m, n) = k \frac{\Omega^{\text{bind}}(m-1, n)}{\Omega^{\text{bind}}(m, n)} \equiv k \times \text{Pr} \left\{ \begin{array}{l} \text{no SSB} \\ \text{at fork} \end{array} \right\}$$

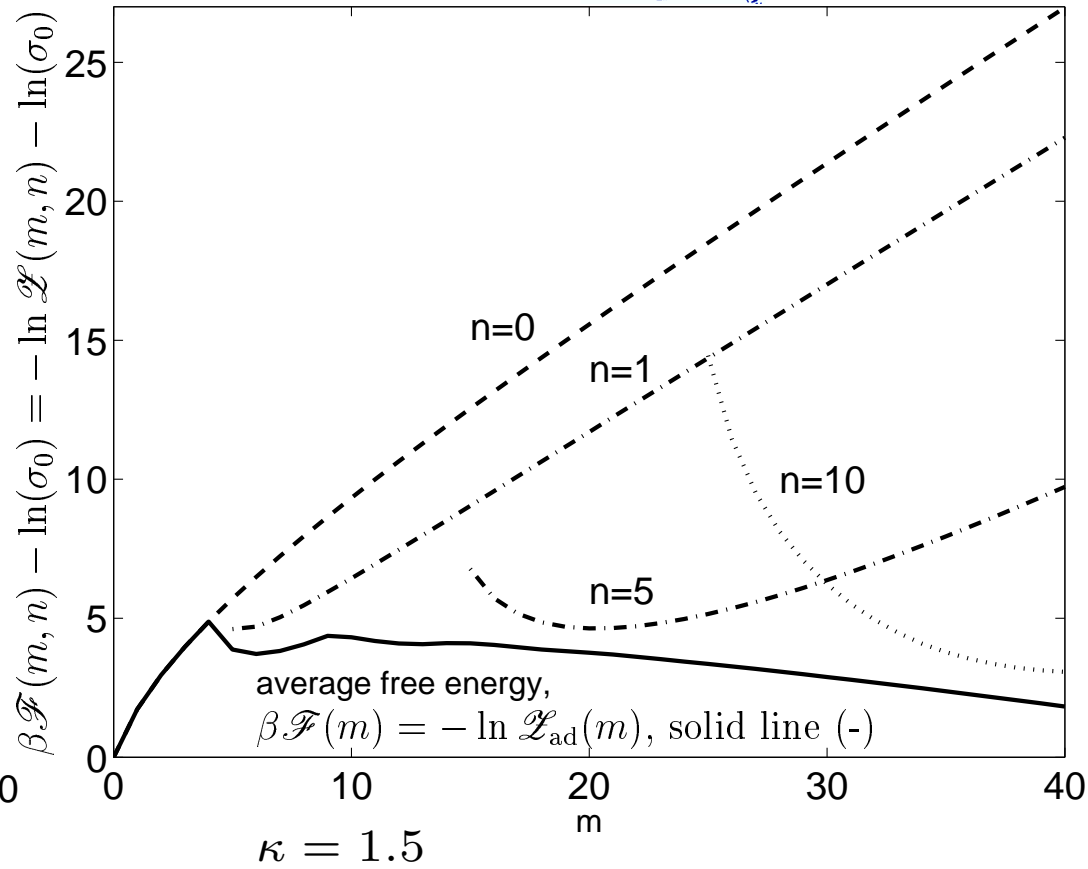
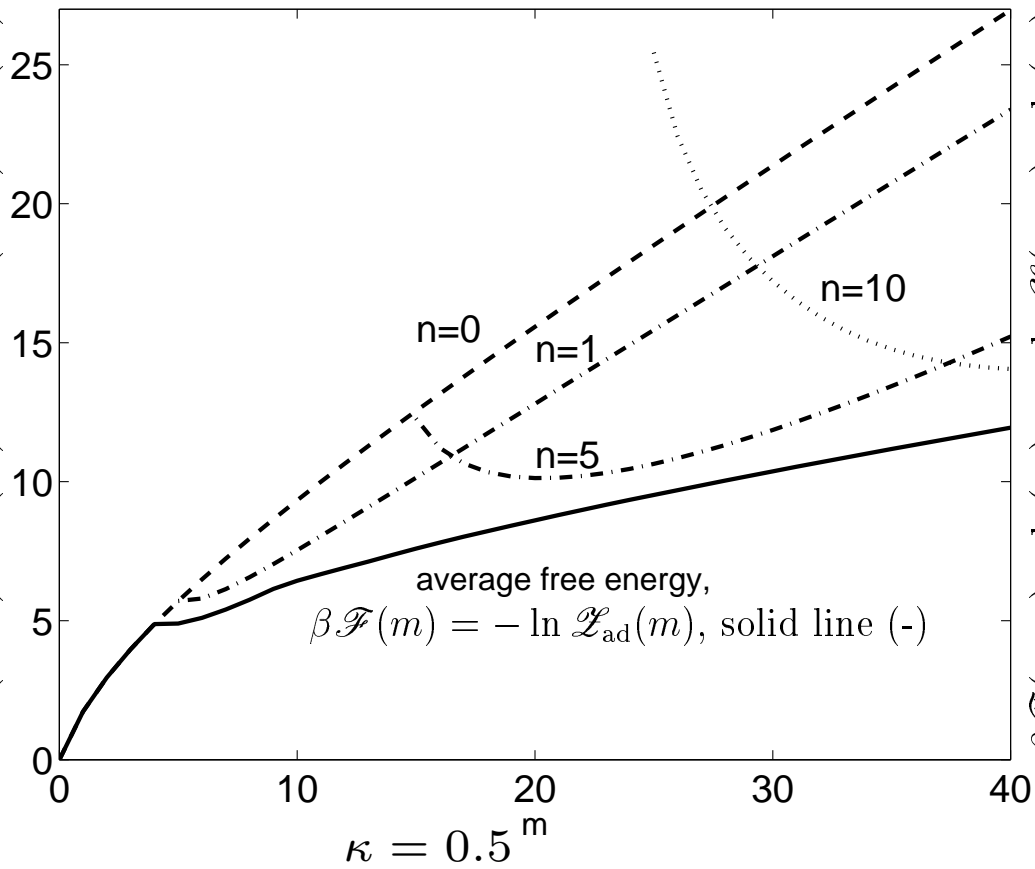
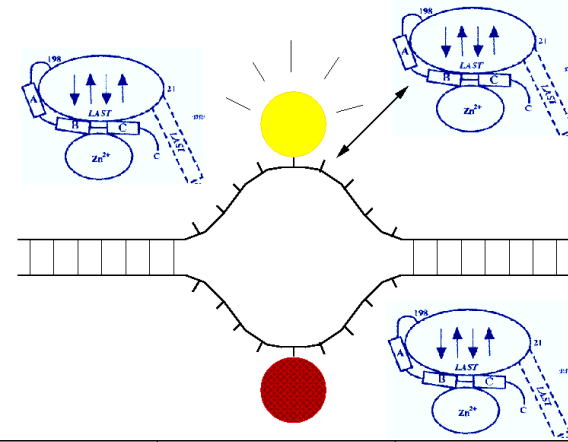
$$r^+(m, n) = \gamma k \kappa (n+1) \frac{\Omega^{\text{bind}}(m, n+1)}{\Omega^{\text{bind}}(m, n)} \equiv \gamma k \kappa \times \text{Pr} \left\{ \begin{array}{l} \exists \text{ slot for} \\ \text{additional SSB} \end{array} \right\}$$

$$r^-(m, n) = n \gamma k$$

$$\gamma \equiv \frac{\{\text{Protein unbinding rate}\}}{\{\text{Bubble zipping rate}\}} = \frac{q}{k}$$

# Breathing bubbles and ssDNA binding proteins (SSBs)

$$\frac{\partial}{\partial t} P(m, n, t) = \mathcal{M}(m, n) P(m, n, t)$$



# DNA bubble dynamics as quantum Coulomb problem

Continuum form of the Poland-Scheraga free energy:

$$\mathcal{F} = \gamma_0 + \gamma_1 \left(1 - \frac{T}{T_m}\right) x + c \ln x$$

Langevin equation for bubble breathing

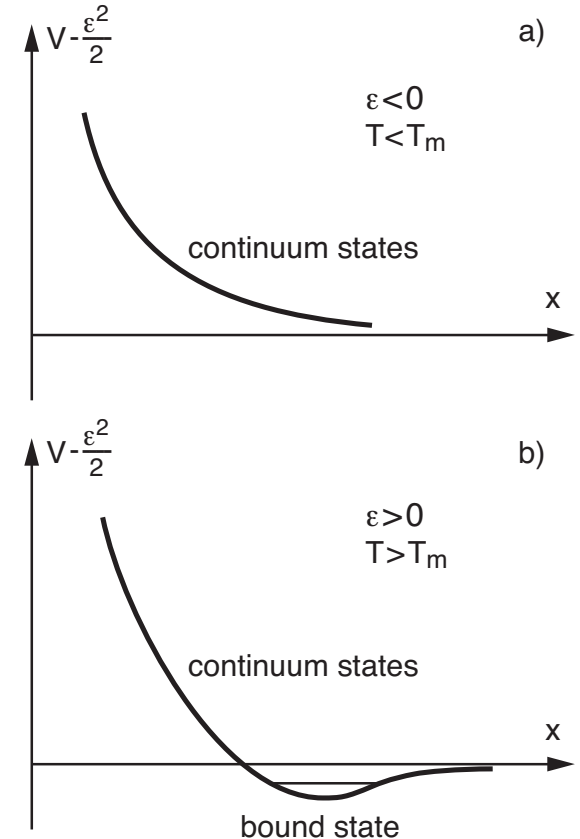
$$\frac{dx}{dt} = -D \frac{d\mathcal{F}}{dx} + \xi(t), \quad \langle \xi(t)\xi(t') \rangle = 2Dk_B T \delta(t - t')$$

Fokker-Planck equation ( $\mu = c/2k_B T$ ):

$$\frac{\partial P}{\partial t} = \frac{\partial}{\partial x} \left( \frac{\mu}{x} - \underbrace{\frac{\gamma_1}{2k_B T} \left[ \frac{T}{T_m} - 1 \right]}_{\epsilon} \right) P + \frac{1}{2} \frac{\partial^2 P}{\partial x^2}$$

With  $P = e^{\epsilon x} x^{-\mu} \tilde{P}$ , obtain imaginary time Schrödinger Eq:

$$-\frac{\partial \tilde{P}}{\partial t} = -\frac{1}{2} \frac{\partial^2 \tilde{P}}{\partial x^2} + \left( \frac{\mu(\mu + 1)}{2x^2} - \frac{\mu\epsilon}{x} + \frac{\epsilon^2}{2} \right) \tilde{P}$$



## Bubble lifetime distribution:

(i)  $T < T_m$ : asymptotically,

$$\wp(t) \simeq x_0^{1+c} e^{|\epsilon|x_0} e^{-\epsilon^2 t/2} t^{-3/2-c/2}$$

$$\mathcal{T} = \int_0^\infty t \wp(t) dt \simeq \frac{x_0 K_{(c-1)/2}(x_0|\epsilon|)}{|\epsilon| K_{(c+1)/2}(x_0|\epsilon|)}$$

(ii)  $T = T_m$ : exact,

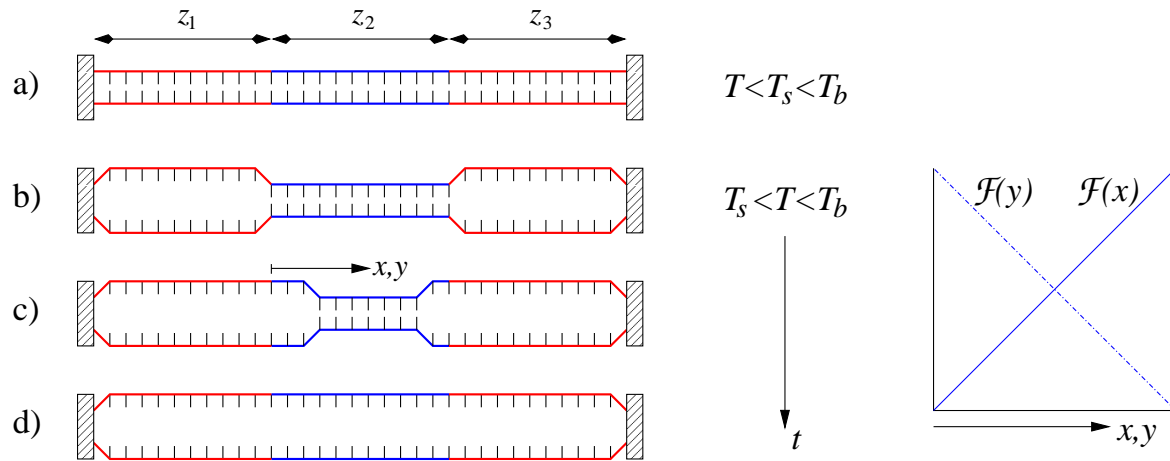
$$\wp(t) = \frac{2x_0^{1+c}}{\Gamma(1/2 + c/2)} e^{-x_0^2/2t} (2t)^{-3/2-c/2}$$

$$\mathcal{T} = \frac{x_0^2}{c-1}, \quad \forall c > 1$$

(iii)  $T > T_m$ : denaturation, bubble coalescence, grand ensemble



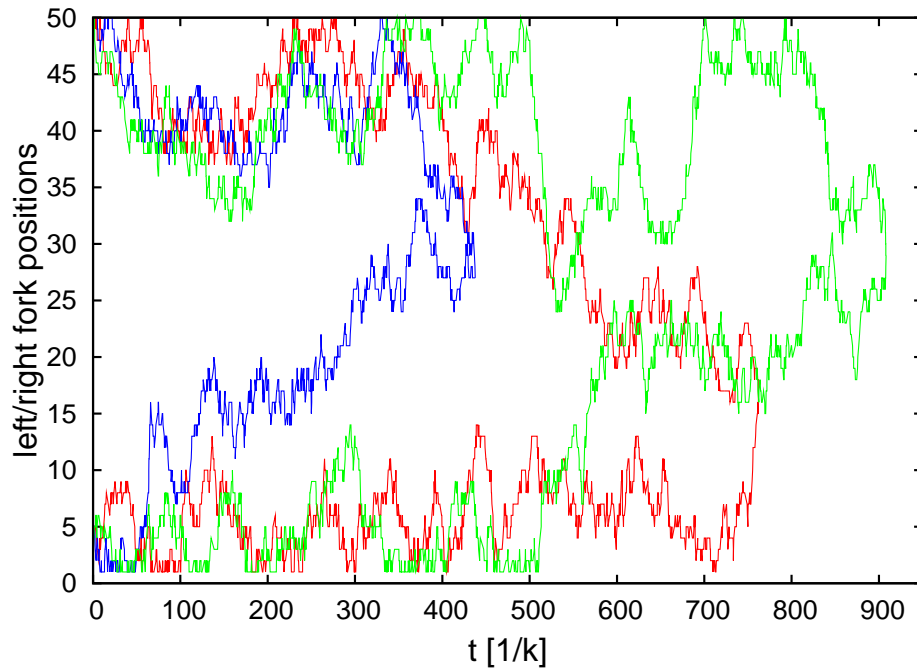
# Bubble coalescence in breathing DNA



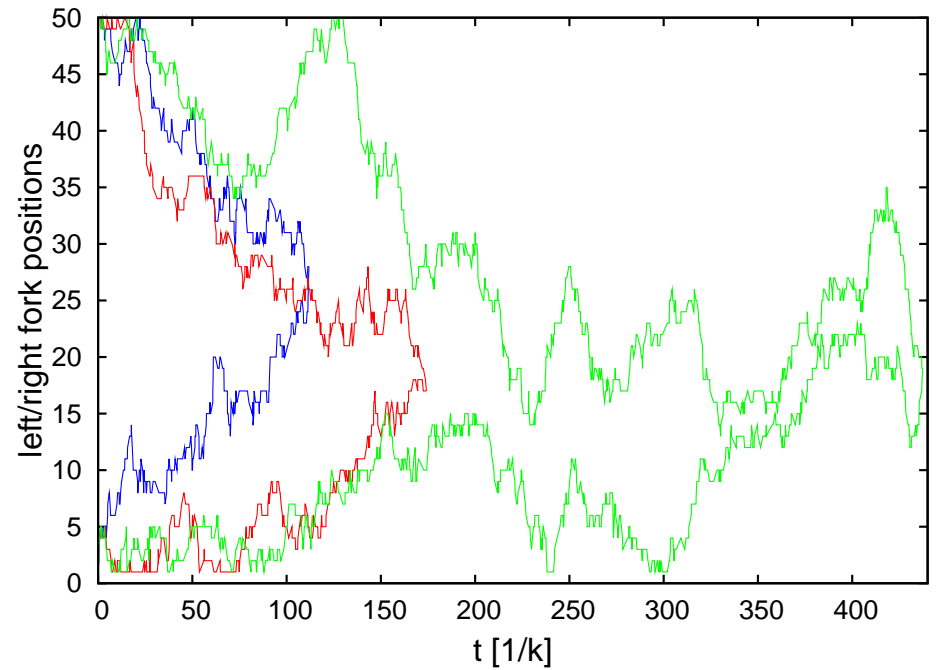
The problem of two viscous walkers in opposite potentials:

$$\frac{\partial}{\partial t} P(x, y, t) = \left( D \left[ \frac{\partial^2}{\partial x^2} + \frac{\partial^2}{\partial y^2} \right] - F \frac{\partial}{\partial x} + F \frac{\partial}{\partial y} \right) P(x, y, t)$$

# Zippering fork trajectories



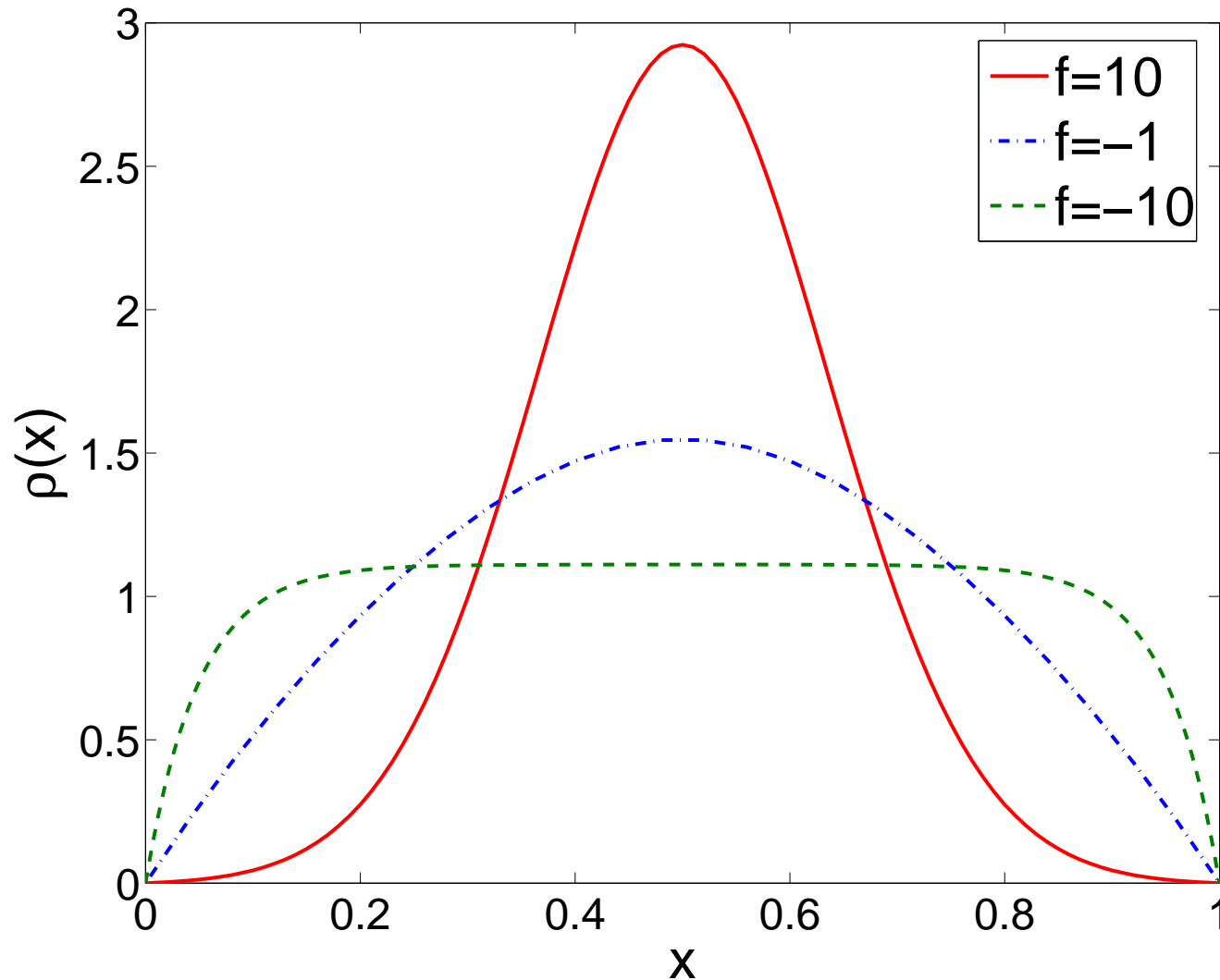
$$u = 0.98$$



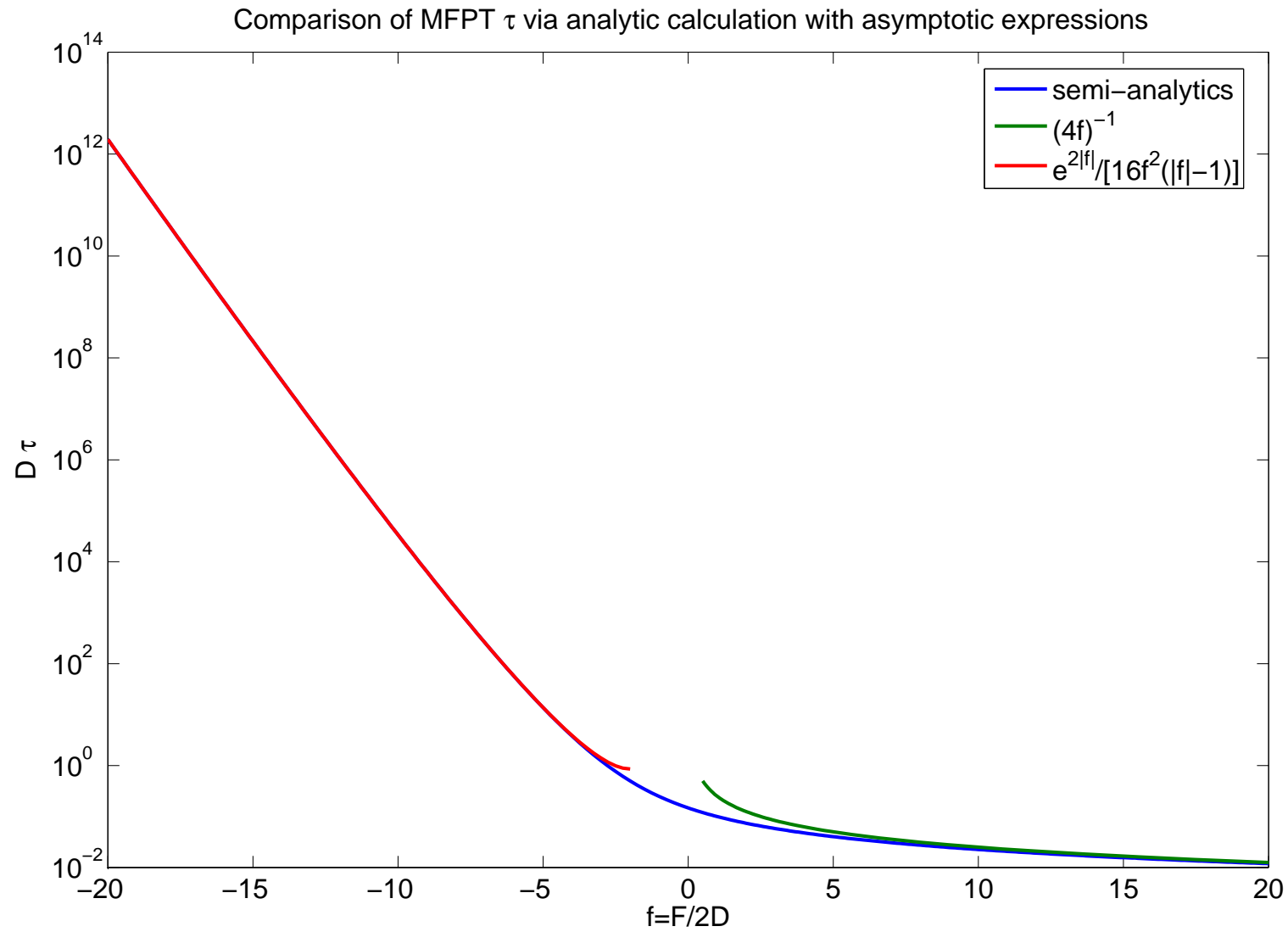
$$u = 1.10$$

## Meeting position:

$f \ll -1$ : Pr to be @  $x$  is  $\propto \exp(-\beta\phi(x))$  with free energy  $\phi(x) = -\int^x f(x')dx' \implies$  joint Pr @  $x$ :  
 $\exp(-\beta[\phi_L(x) + \phi_R(x)]) \approx \text{const}$



# Mean coalescence time



# Coalescence time density, comparison of methods

